ACS Chemical Neuroscience

Protective Effect of Natural Products against Huntington's Disease: An Overview of Scientific Evidence and Understanding Their Mechanism of Action

Pei Teng Lum, Mahendran Sekar,* Siew Hua Gan, Srinivasa Reddy Bonam, and Mohd. Farooq Shaikh



dysfunction. To the best of our knowledge, there is no treatment available to completely mitigate the progression of HD. Among various therapeutic approaches, exhaustive literature reports have confirmed the medicinal benefits of natural products in HD experimental models. Building on this information, this review presents a brief overview of the neuroprotective mechanism(s) of natural products against *in vitro/in vivo* models of HD. Relevant studies were identified from several scientific databases, including PubMed, ScienceDirect, Scopus, and Google Scholar. After screening through literature from 2005 to the present, a total of 14 medicinal plant species and 30 naturally isolated compounds investigated against HD



based on either *in vitro* or *in vivo* models were included in the present review. Behavioral outcomes in the HD *in vivo* model showed that natural compounds significantly attenuated 3-nitropropionic acid (3-NP) induced memory loss and motor incoordination. The biochemical alteration has been markedly alleviated with reduced lipid peroxidation, increased endogenous enzymatic antioxidants, reduced acetylcholinesterase activity, and increased mitochondrial energy production. Interestingly, following treatment with certain natural products, 3-NP-induced damage in the striatum was ameliorated, as seen histologically. Overall, natural products afforded varying degrees of neuroprotection in preclinical studies of HD via antioxidant and anti-inflammatory properties, preservation of mitochondrial function, inhibition of apoptosis, and induction of autophagy.

KEYWORDS: Huntington's disease, natural products, neuroprotective, neurodegenerative, 3-nitropropionic acid, herbal medicine

INTRODUCTION

Huntington's disease (HD) is an autosomal dominant, inherited neurological disorder associated with a pathogenic expansion of cytosine-adenine-guanine (CAG) trinucleotide repeats in exon 1 of the huntingtin gene (Htt).¹ The onset of HD occurs between 30 and 50 years (mean survival, 15–20 years). HD occurring before 21 years of age is deemed as a juvenile HD, while the other extreme, "late-onset", is that occurring after 60 years old.²

The data from meta-analyses have reported a global prevalence of approximately 2.7 in 100 000 HD with the lowest incidence seen among Asians and the highest in Western populations.³ However, the overall incidence of HD worldwide remains unclear because epidemiological evidence from Asia and African populations to date are limited to only clinical reviews and case studies.⁴ Despite discerning common HD genetic origins, disease progression tends to vary based on an individual's mutation rates, diagnostic stigma, and criteria.

Mutant Htt (mHtt), a mutated functional protein carrying abnormal and elongated polyglutamine (polyQ) is the main factor in contributing to the development of HD, exhibiting putative detrimental properties to the entire parts of the brain.² Pathologically, a toxic gain function of mHtt tends to be devasting to the intracellular pathways in altering proteostasis and protein degradation following transcription and synaptic dysfunction, striatal excitotoxicity, dopamine toxicity, mitochondrial dysfunction, metabolic impairment, oxidative stress, and neuronal cell death.⁵ After a premanifest period of HD, its clinical characteristics are diagnosed based on behavioral components, including cognitive and motor signs and symptoms. Paradoxically, there are merely symptomatic treatments often used in ameliorating the negative impacts of

Received: December 28, 2020 Accepted: January 11, 2021

In the second se

A



Figure 1. Pathophysiology of HD. Mutation of *Htt* characterized with repeat expansion of CAG trinucleotides is the key factor in HD. Abnormal aggregation of mutant Htt protein may cause toxic effects in neurons, leading to a series of pathogenic mechanisms associated the alteration in proteostasis and protein degradation following mitochondrial dysfunction, oxidative stress, transcription and synaptic dysfunction, axonal transport impairment, and a series of metabolic impairments subsequent to neurodegeneration. Abbreviations: HD, Huntington's disease; *Htt*, huntingtin gene; MAPK, mitogen-activated protein kinase; MSN, medium spiny neurons; NF- κ B, nuclear factor kappa light chain enhancer of activated B cells; UPS, ubiquitin proteasome system.



Figure 2. Normal versus mutated huntingtin. Abbreviations: Htt, huntingtin; mHtt, mutant huntingtin; wtHtt, wild-type huntingtin.

HD. Therefore, researchers are oriented toward implementing therapeutic strategies by targeting the pathway of mHtt production to address the cause of HD pathophysiology.

Nevertheless, in defiance to the existing therapeutic approaches, HD impact is still not fully controlled.

To overcome the above-mentioned concerns, investigation has been devoted to the isolation of novel compounds from a variety of natural products in modulating relevant neurodegenerative disorders. Up until then, a plethora of traditional treatments based on natural products have been shown to possess a wide range of therapeutic benefits for HD under in vitro and in vivo models. Indeed, based on relevant studies, natural products offer neuroprotection in experimental models predominantly through the antioxidant defense system, scavenging free radicals, neutralization of reactive oxygen species (ROS), reduction of oxidative stress, preservation of mitochondrial function, anti-inflammatory protection, inhibition of apoptosis, and induction of autophagy. To gain insight into the potential role of natural products, an overview of various natural products against HD in preclinical studies is presented in this review. We attempt to outline the pathophysiology of HD, mapping the clinical manifestations and current therapeutic approaches for HD. Subsequently, a summary of information gathering the neuroprotective potency of natural products against HD animal models is presented. Possible neuroprotective mechanisms are also highlighted and discussed based on the relevant findings from HD in vitro and in vivo models.

HUNTINGTON'S DISEASE

Pathophysiology of HD. The pathophysiology of HD is presented in Figure 1. Huntingtin (Htt) is a protein formed by more than 3100 amino acids, encoded by a gene located at chromosome 4.² Underlying the molecular mechanism, mutation of Htt gene characterized by the repeat expansion of CAG trinucleotides within exon 1 is the major factor in the pathophysiology of HD⁶ (Figure 2). The number of the CAG repeats is predominantly chosen as the primary determinant of HD severity due to its instability and change in length with either a decrease of 1-2 units or an increase of 1-4 units during parental transmission.^{2,6} For the normal population, the repeat is said to be polymorphic in the range of 6-35 units. Clinically, an individual with CAG repeats of 36-39 units has HD with incomplete penetrance correlated with later onset. They live a normal lifespan with no symptoms apparent for diagnosis. However, when the CAG repeats expand to above 40 units, it is considered as the longest repeats with high penetrance triggering signs of early onset diagnostic for HD. The stretched tract of polyQ at the amino terminus of the translated Htt protein is linked with the abnormal aggregation of protein and a complex loss of function phenotype.

The expanded polyQ tract with a high order of amyloid fibers and insoluble β -pleated sheets is prone to aggregation that can disrupt the intracellular function directly by affecting proteostasis and impairing protein degradation, thus resulting in transcription dysregulation.^{2,7} The formation of protein aggregates in HD is a complex process encompassing sequestering of other proteins into the mHtt aggregates. Based on the said hypothesis, misfolded Htt cannot bind and inhibit the activation of caspase-3. N-terminal fragments produced from the caspase-3-mediated proteolysis of mHtt tend to induce proteolytic cleavage and additional protease aggregation. Interestingly, proteasome components and heat shock proteins (HSPs) may form aggregates of mHtt. Subsequently, the functional proteasomes and heat shock protein 70 (Hsp70) in neurons are progressively depleted in vitro, leading to a proteostasis collapse, accompanied by the chronic expression of mHtt and increased protein aggregation.⁸

On the other hand, imbalance of the ubiquitin-proteasome system (UPS), which occurs due to the overwhelming proteostasis induced by mHtt, was observed in the brain tissue of mouse models and HD patients, thus suggesting that impaired protein degradation pathways may be involved.⁵

Transcriptional dysregulation has been reported to be linked with progressive expression of mHtt in HD. Studies^{2,7} have shown that various transcriptional factors, including specific protein-1 (Sp1), cyclic-adenosine monophosphate (cAMP) response element-binding protein (CREB)-binding protein (CBP), thymidine-adenine-thymidine-adenine (TATA)-binding protein (TBP), p53, and brain-derived neurotrophic factor (BDNF), tend to interact with *mHtt* while expressing mHtt. On the other hand, the level of CBP is decreased parallel to histone hypoacetylation as well as inhibition of CBP-regulated transcription. In addition, Sp1 gene transcription is also inhibited as confirmed by the reduction of Sp1 binding onto specific promoters purported to be due to dissociation of binding of Sp1 from the particular promoter on mHtt. Moreover, suppression of the BDNF gene contributed by mHtt leads to striatal neuron degeneration.

Additionally, it is suggested that neutrophil aggregation caused by mHtt affects axonal transport by physically blocking the signal transport within the axon terminus leading to impairment of neuronal synaptic transmissions.⁹ The expression of pre- and postsynaptic neurotransmitter receptors is also perturbed, contributing to synaptic dysfunction.^{10,11} Besides intracellular dysfunction, various neurodegenerative mechanisms, including cortico-striatal excitotoxicity,¹² mitochondrial dysfunction,¹³ metabolic energy impairment, and oxidative stress,¹⁴ followed by cell death as a result of apoptosis and dysregulated autophagy, have been linked to HD pathophysiology.²

Interestingly, wild-type Htt (wtHtt) contributes to normal embryonic development. However, polyglutamine expansion obstructs the interaction of wtHtt with postsynaptic density 95 (PSD-95), leading to (1) the sensitization of *N*-methyl-Daspartic acid (NDMA) and kynurenic acid (KA) receptors¹² and (2) glutamate-mediated excitotoxicity. mHtt also induces the tyrosine phosphorylation of NDMA receptors, promoting further sensitization.¹⁵ In addition, as confirmed by Tang et al.,¹⁶ the expression of metabotropic glutamate receptor subtype 5 (mGluR5) and NMDA receptor subtype 2B (NR2B) is involved in the mHtt-induced excitotoxicity in striatal medium spiny neurons (MSNs) of yeast artificial chromosome (YAC) transgenic mice.

Collectively, chronic NDMA activation and striatal dopamine (DA) depletion synergistically induce an increase in intracellular calcium levels, causing mitochondrial dysfunction.¹⁶ Several studies have supported the fact that mitochondrial dysfunction and metabolic impairment are important pathological hallmarks of HD.^{2,6} In vitro studies indicated the loss of mitochondrial function caused by 3nitropropionic acid (3-NP) via the expression of cytochrome *c* concomitant with caspase activation eventually.¹⁷ The release of cytochrome c triggers caspase activation, causing cleavage of mHtt and its translocation into the nucleus. According to Bae et al.,¹⁹ nuclear mHtt can induce the expression of p53 (a vital transcription factor, which mediates the expression of several mitochondrial proteins, including Bax) in the primary neuronal cultures from mouse embryos, leading to mitochondrial abnormalities. Additionally, it has been suggested that mHtt fragments can associate with the



Figure 3. Interaction of mutant Htt with mitochondrial protein and subsequent pathogenic changes in HD neurons. Abbreviations: ATP, adenosine triphosphate; Drp1, dynamin-1-like protein; GSH, reduced glutathione; HD, Huntington's disease; Htt, huntingtin; ROS, reactive oxygen species; SOD, superoxide dismutase.

outer mitochondrial membrane by causing the opening of mitochondrial permeability transition pores and disrupting the electron transport chain (ETC).¹⁹

Two important but opposing forces that maintain mitochondrial shape and structure are mitochondrial fission and mitochondrial fusion. The plethora of evidence suggests that mitochondrial dynamics become disturbed in neurodegenerative diseases like HD. In the aged neurons that express mutant proteins, such as mutant Htt, an imbalance between fission and fusion leads to abnormalities in mitochondrial structure and function and neuronal damage. Figure 3 shows mutant huntingtin interaction with the mitochondrial protein dynamin-1-like protein (Drp1) and subsequent pathogenic changes in HD neurons. Overall, available evidence suggests that Drp1 intermingles with mutant Htt and increases Drp1 enzymatic activity in HD-affected regions, leading to both synaptic and neuronal damage. The mitochondrial dysfunction is the key that leads to oxidative stress and hence low synaptic ATP. In connection to this, the increased oxidative stress activates inducible nitric oxide synthase (iNOS) expression and inhibits the endogenous enzymatic antioxidants, including superoxide dismutase (SOD), glutathione (GSH) peroxidase, and cAMP, all of which lead to protein and lipid peroxidation as well as DNA mutation. Subsequently, this leads to neuronal damage and loss, which are key hallmarks of HD.

Clinical Manifestation of HD. The course of the life of a person with one parent with HD can be divided into at-risk, preclinical, and clinical stages.²⁰ The presence of elongated CAG repeats indicates that the individual is in the "at-risk" stage. On the other hand, the presence of a *mHtt* gene, predisposes an individual to progress to the preclinical followed by the clinical stages. The clinical features of HD predominantly comprise behavioral components, including cognitive and motor signs and symptoms. In the early stage, psychiatric disturbance is the most frequent symptom, appearing before the onset of the motor symptoms. Never-

theless, behavioral symptoms vary between different patients although the signs and symptoms commonly negatively impact daily functioning and relationships.

Among the various psychiatric symptoms, irritability is deemed as the first. Retrospectively, the symptoms may also occur in any of the stages of HD. Irritability is expressed in different forms, including showing severe conflict with others or worse, physical aggression. Both irritability and aggression are usually caused by compulsion and obsession with an individual's lifestyle. Commonly, depression includes anxiety, and guilt and low self-esteem are present throughout the different HD stages. Apathy is difficult to distinguish from depression. Nevertheless, HD patients with apathy commonly express passive behavior in lieu of a lack of interest. As the disease progresses, psychosis may appear concomitant with a cognitive decline.²⁰

Significant cognitive changes are one of the noticeable signs of HD and exist long before the inception of the first motor symptoms. Particularly, a patient's memory is impaired although language ability is comparatively spared. Subsequently, psychomotor processes tend to be more severely retarded. Additionally, cognitive decline is also associated with a series of unique functions, including an inability to plan or organize life single-handedly due to loss of mental agility. Owing to this fact, patients are incapable of making good mental judgments.

Motor incoordination is involuntary. Initially, the unwanted movements tend to affect the distal extremities, including the fingers and the toes as well as small facial muscles. Gradually, the defect spreads to the muscles, starting from the distal area to more and more proximal and even toward axial, thus making daily walking gait unstable and a challenge. In fact, dystonia, characterized by a slower movement accompanied by an increased tone of muscles and culminating in abnormal postures, is the first motor symptom in HD.²¹

Among all the stated motor symptoms, choreatic movements, particularly continuous movements of the eyes,



Figure 4. Medicinal plants with potential against HD in experimental models.

Fable 1. List of Natural Products	(Medicinal Plants) with Potentia	l against HD in I	Experimental Models
-----------------------------------	-------------------	-----------------	-------------------	---------------------

natu	ural plants				
botanical name	common name	family name	part of plants	natural compounds	refs
Anemarrhenae asphodeloides		Asparagaceae	rhizome	mangiferin	Piwowar et al. ³³
Calendula officinalis	marigold	Asteraceae	flower		Shivasharan et al. ⁴⁵
Celastrus paniculatus	jyotishmati, malkangni or kangani	Celastraceae	seed		Malik et al. ⁴⁷
Centella asiatica	Gotu Kola	Umbelliferae	leaf		Shinomol et al. ⁵⁰
Convolvulus pluricaulis	Shankhpushpi	Convolvulaceae	whole plant	scopoletin	Kaur et al. ⁵⁴
Ficus religiosa	pimpala or pipal tree	Moraceae	leaf		Bhangale et al. ⁵⁸
Ginseng radix	Korean red ginseng	Araliaceae	root	ginsenosides	Jang et al. ⁶⁰
Luehea divaricata	acoita-cavalo	Tiliaceae	leaf		Courtes et al. ⁶³
Panax quinquefolius	American ginseng	Araliaceae	leaf and stem	panaxadiols (Rb ₁ , Rb ₃ , Rd)	Lian et al. ⁶⁵
Phoenix dactylifera	date palm	Arecaceae	fruits		Essa et al. ³⁶
Psoralea corylifolia		Leguminosae	seed		Im et al. ³⁹
Punica granatum	pomegranate	Punicaceae			Al-Sabahi et al. ⁴³
Withania somnifera	Ashwagandha	Solanaceae	root		Kumar and Kumar ⁶⁹
Zingiber officinale	ginger	Zingiberaceae	root		Sharma et al. ⁷¹

eyebrows, head, and tongue, exist consistently during the disease progression. Eventually, all HD patients will develop some forms of hypokinesia, slowly leading to bradykinesia (slowness of movement), followed by akinesia (difficulty in initiating movement). These altered motor performances progressively trigger an individual to have difficulties in standing and walking. Talking and swallowing problems are other more prominent motor symptoms for HD.²⁰ Having said that, the time when the signs and symptoms start to interfere with daily tasks are hugely dependent on the type of daily activities and work of HD patients.

Current Therapies in HD. To date, there are no promising treatments for the long-term unwanted effects of HD, which are being combated by symptomatic prevention and treatments for mitigating the psychiatric, cognitive, and motor deformities of HD. In the recent status of HD drug therapies, only tetrabenazine is approved by Food and Drug Administration

(FDA) in the treatment of chorea and other HD-related motor symptoms.^{22–24} Antipsychotic drugs such as tiapridal and olanzapine are believed to have the potential to treat chorea by modulating dopamine receptors. Their performances are currently being evaluated under a phase III clinical trial.^{23,25}

Given the paucity of proven potency of existing symptomatic treatments for HD, the current targets of emerging treatments are focused on the development of mechanism-based therapies in parallel with the concepts and knowledge of possible pathways involved in HD pathogenesis. Therefore, growing therapeutic strategies have been aimed at Htt lowering, Htt modulation, immunomodulation, synaptic modulation, and stem cell transplantion approaches, which may be either RNA-or DNA-based.³ In the former, antisense oligonucleotides (ASOs), synthetic single-stranded DNA sequences, and RNA interference (RNAi) can induce advanced degradation of mHtt to lower mHtt levels.²⁶ Normally, two types of ASOs are used

Review

Table 2. Summary of Natural Products (Medicinal Plants) with Potential against HD as Confirmed by In Vitro Studies^a

natural plants (botanical name)	dose	IC ₅₀	EC ₅₀	cell lines	model	molecular outcomes	mode of action	refs
Anemarrhena asphodeloides	0.5, 1 μ g/mL ^b	С	С	PC-12 cells	3-NP	cell viability ↑	antioxidant system	Piwowar et al. ³³
Phoenix dactylifera	$1-1000 \ \mu g/mL$	276.4 µg/mL	С	PC-12 cells	3-NP	cell viability ↑, ATP ↑, LDH activity ↓, ROS ↓, MDA ↓, NO ↓, SOD ↑, GPx ↑, GSH ↑	scavenge ROS	Essa et al. ³⁶
Psoralea corylifolia	10, 50, 100 μg/ mL ^b	с	С	PC-12 cells	3-NP	cell viability ↑, ATP ↑, OCR ↑, mMP ↑; mitochondrial superoxide ↓	restoration of mitochondrial function	Im et al. ³⁹
Punica granatum	40 µM	С	С	PC-12 cells	genetically modified	ROS ↓, RNS ↓, MDA ↓, LDH activity ↓, NO ↓, LPO ↓	neutralize ROS, ↑ antioxidant gene expression	Al-Sabahi et al. ⁴³

^{*a*}Abbreviations: 3-NP, 3-nitropropionic acid; A_{2A}R, adenosine A_{2A} receptor; AMPK, AMP-activated protein kinase; Atg, autophagy; ATP, adenosine triphosphate; CGNs, cerebellar granule neurons; LC3-I, cytosolic form of LC3; LC3-II, LC3-phosphatidylethanolamine conjugate; LDH, lactate dehydrogenase; LPO, lipid peroxidation; MDA, malondialdehyde; MEF, mouse embryonic fibroblasts; mHtt, mutant huntingtin; mMP, mitochondrial membrane potential; MSN, medium spiny neurons; mTOR, mammalian target of rapamycin; NMDA, *N*-methyl-D-aspartate; NO, nitric oxide; OCR, oxygen consumption rate; PC-12, pheochromocytoma-12; PKA, protein kinase; RNS, reactive nitrogen species; ROS, reactive oxygen species; WT, wild-type. ^{*b*}Concentration dependent. ^{*c*}Not determined.

(1) allele-specific ASOs, which target only *mHtt*, and (2) allele nonspecific ASOs targeting *wtHtt* and *mHtt*. Nevertheless, reduced levels of *wtHtt* and *mHtt* may confer potential risks of deleterious effects, making these therapeutic approaches not well-established. Currently, IONIS-*HTT*_{Rx}(RG6042), one of the allele nonspecific ASOs, is being tested for its safety in phase III clinical stages of development.²⁶

On the other hand, the mHtt gene with elongated CAG repeats can be targeted by using (1) zinc finger motif proteins (ZFPs) to repress its transcription or (2) clustered regularly interspaced short palindromic repeats (CRISPR) with CRISPR-associated system 9 (CRISPR-Cas9), which is a genome editing tool to edit mutated *Htt* DNA sequences.²⁷ To date, RNAi, ZFPs, and CRISPR-Cas9 are still in preclinical stages. Another useful alternative strategy is to modulate Htt homeostasis by inhibiting its aggregation or promoting its clearance. In fact, immunotherapy is likely to be a novel and promising approach to modulate the status of pro-inflammatory profile in HD patients. Despite negative findings from other disease-modifying clinical trials, immunotherapies via targeting semaphoring 4AD (SEMA4D) for modulation of neuroinflammation pathways still yield promising findings.²⁸ Finally, depending on an individual's disease progression, the development of surgical treatment such as stem cell transplantation has also been suggested.

RESULTS AND DISCUSSION

Protective Effects of Natural Products (Medicinal Plants) against HD. The neuroprotective effect of natural products in HD experimental models has been extensively studied. Hence, a variety of medicinal plants (Figure 4) that have been studied in preclinical models of HD is listed in Table 1, while a comprehensive detail of their neuroprotective effects against HD under *in vitro* and *in vivo* models are summarized in Tables 2 and 3, respectively.

In Vitro Studies. Anemarrhenae asphodeloides Bunge. The rhizomes of Anemarrhenae asphodeloides Bunge, which belongs to the Asparagaceae family, are a widely used traditional medicine in Eastern Asian countries such as China, Japan, Korea, and Mongolia. The rhizomes of Anemarrhenae asphodeloides are also well-known as Zhimu and Yanghuzi in Chinese medicine and Chimoi and Jimo in Japanese and Korean medicine, respectively.²⁹ According to Chinese medicine, Anemarrhena rhizome is recognized as a traditional warm herb for yin nourishing, heat-clearing, and kidney, lung, and stomach meridian entering correlated with the curative function of treating dry cough, fevers, night sweats, menopause syndrome, and diabetes.^{29,30} It contains an abundance of active constituents, such as xanthones, saponins, alkaloids, flavonoids, anthraquinones, phenylpropanoids, and organic acids^{31,32} with a series of pharmacological benefits, including anti-inflammatory, antibacterial, antipyretic, antiviral, antidiabetic, and anticoagulation.^{29,33} A recent study was conducted by Piwowar et al.³³ to evaluate the potential protective effects of the Anemarrhena rhizome ethanolic extract toward a HD in vitro model with 3-NP induced neurotoxicity in pheochromocytoma (PC-12) cells. Suppression of 3-NP induced cytotoxic activity and enhancement of cell proliferation were observed in Anemarrhena rhizome extract-treated cells in which cell apoptosis and morphological changes were prevented. However, the protective action was dependent on the method of treatment, concentration, and incubation time. The finding reports that the xanthone fraction of ethanolic Anemarrhena rhizome extract (0.5 and 1.0 μ g/mL) offered the protection for preincubated PC-12 cells in a concentrationdependent manner.

Phoenix dactylifera Linn. Phoenix dactylifera Linn (Arecaceae) is also known as date palm, a well-known woody fruit tree predominantly cultivated in Middle, Eastern, and Western Asia and North Africa. Date palm fruits are highly nutritious and rich in carotenoids, sterols, flavonoids, lignans, and phenolic acid, which are potent antioxidants.³⁴ They are edible fruits with high carbohydrates and lower tannins and moisture content. Pharmacological activities of date palm fruits including antiviral, anti-inflammatory, antifungal, anticancer, nephroprotective, hepatoprotective, and antihyperlipidemic properties have been extensively reported in previous studies.^{35,36} Recently, it was reported to possess neuroprotective effects in a 3-NP induced HD in vitro model.³⁶ In the study, the ethanolic extract of date palm fruits was observed to ameliorate oxidative stress-induced mitochondrial dysfunction evidenced by restored intracellular ATP production in 3-NP intoxicated PC-12 cells. It was attributed to the antioxidant properties of the date palm fruits in reducing the ROS generation accompanied by an increase of endogenous enzymatic antioxidants (SOD and GPx) and nonenzymatic antioxidants such as reduced glutathione (GSH).³⁶ In addition, the results from cytotoxicity assay demonstrated that the

Studies ^a
Vivo
In
bу
Confirmed
Ę
against I
Potential
with
Plants)
(Medicinal
Products
of Natural
Summary o
÷

Table

						outcomes			
natural plants (botanical	dose (mg/kg);	duration	animal mod-						
name)	route	(days)	els, sex	toxin	behavioral	biochemical	histopathological	mode of action	refs
Calendula of- ficinalis	100, 200; Po	5	Wistar rats, female	3-NP, ip	<pre>locomotor count ↑, TL ↓, body balance ↑, hind limb function ↑, grip strength ↑</pre>	LPO ↓; GSH †; TSH †; GST †; CAT activity †; NIT ↓	striatal degeneration \downarrow	antioxidant, anti-inflam- matory, and estrogenic protection	Shivasharan et al. ⁴⁵
Celastrus pan- iculatus	100 ^b , 200; Po	20	Wistar rats, male	3-NP, ip	impairment in grip \downarrow locomotor activity \uparrow TL \downarrow , TSTQ \uparrow	GSH †, SOD †, CAT activity †; MDA ↓, NIT ↓		↑ antioxidant defense system, ↓ glutamate toxicity	Malik et al. ⁴⁷
Centella asiat- ica	5, po	10	prepubertal mice, male	3-NP, ip		ROS 4, MDA 4; protein carbonyl 4; GSH 1, TSH 1; SOD 1, GST \uparrow , GPx 1; MTT 1, CS 1; SDH activity 1, MDH 1; ETC enzymes activities 1; Na ⁺ , K ⁺ and ATPase \uparrow		antioxidant defense and preservation of mito- chondrial integrity	Shinomol et al. ⁵⁰
Convolvulus pluricaulis	10, 20 ^b ; po	14	Wistar rats, male	3-NP, ip	locomotor, grip strength, and rotarod activity †	GSH †, SOD †; MDA ↓, NIT ↓		the antioxidant defense system system	Kaur et al. ⁵⁴
	100, 200 ^b ; Po	20	Wistar rats, male	3-NP, ip	locomotor and rotarod activity \uparrow , TL \downarrow	GSH †, SOD †, CAT activity †; MDA ↓, NIT ↓		↑ antioxidant defense system	Malik et al. ⁵²
Ficus religiosa	100, 200, 400 ^b ; Po	14	Wistar rats, male	3-NP, ip	locomotor and muscle activity ↑	GSH †, SOD †, CAT activity †; AChE ↓; MDA ↓	swelling of cells \downarrow , density of cells \uparrow , neuronal damage \downarrow	↓ oxidative stress	Bhangale et al. ⁵⁸
Ginseng radix	50, 100, 250; ^c Po	18	ICR mice, male	3-NP, ip		microglial activation 1, expression of TNF-a, IL-1 β , IL-6 and iNOS 4, activation of JNK, ERK 4, MAPKs and NF-xB 4	lesion volume and striatal degeneration ↓	inhibition of MAPKs and NF-xB	Jang et al. ⁶⁰
Luehea divari- cata	500, 1000; ig	10	Wistar rats, male	3-NP, ip	rotarod latency ↓	ROS 4; TBARS 4; AChE 4; GSH/GSSG ratio restored		antioxidant defense sys- tem	Courtes et al. ⁶³
Panax quin- quefolius	10, po	S	SD rats, male	3-NP, ip	behavioral change and motor impairment ↓		loss of neurons in the hippocampus \downarrow , striatal lesion \downarrow	scavenging free radicals	Lian et al. ⁶⁵
					hypothermia ↓, PPI deficit ↓, locomotor activity ↑	GABA 1, DA 1, S-HT 1, NE 1; GSH 1, MDA 1	degree of pathological injuries in brain \downarrow	antioxidant defense sys- tem	Mahdy et al. ¹¹³
Withania som- nifera	100, 200; ^c Po	14	Wistar rats, male	3-NP, ip	motor activity $\uparrow,$ grip strength \uparrow	MDA J, NIT J; SOD ↑, CAT activity ↑; LDH activity ↓; NADH ↑, SDH activity ↑, MTT ↑		antioxidant defense sys- tem	Kumar and Kumar ⁶⁹
Zingiber offici- nale	100, 200; ^c Po	7	Wistar rats	3-NP, ip	locomotor activity ↑, grip strength ↑, memory performance ↑	AchE J; MDA J; GSH ↑	brain inflammation 4, necrosis 4, gliosis 4	anticholinesterase po- tency	Sharma et al. ⁷¹
^a Abbreviation associated X 1	is: 3-NP, 3 vrotein: Bcl	-nitroprol	pionic acid; 5-	-HT, 5- RDNF	hydroxytryptamine; A _{2A} R, ad	enosine A _{2A} receptor; AC, adenyl cyclase; AChE, acetyc	holinesterase; ATP,	adenosine triphosphate	; Bax, Bcl-2-

enhancer of activated B cells, NQO1, NAD(P)H:quinone oxidoreductase 1; NrP, nuclear factor erythroid 2-related factor 2; po, per os, oral administration; PPI, prepulse inhibition; ROS, reactive oxygen species; sc, subcutaneous injection; SD, Sprague–Dawley; SDH, succinate dehydrogenase; SOD, superoxide dismutase; TBARS, thiobarbituric acid reactive substances; TH, tyrosine hydroxylase; TL, transfer latency; TNF-*a*, tumor necrosis factor *a*; TSH, total thiols; TST, tail suspension test: TSTO. time snent in target market administration and environments.

HO1, heme oxygenase-1; Hsp70, heat shock protein 70; Htt, huntingtin; ICR, Institute of Cancer Research; ig, intragastric gavage; ip, intraperitoneal injection; IL-1ß, interleukin-1ß; IL-6, interleukin 6; iNOS, inducible nitric oxide synthase; JNK, c-Jun N-terminal kinase; LDH, lactate dehydrogenase; LPO, lipid peroxidation; MAPK, mitogen-activated protein kinase; MDA, malondialdehyde; MDH, malate dehydrogenase; mHtt, mutant huntingtin; NAD, nicotinamide adenine dinucleotide; NADH, NAD + hydrogen (H); NE, norepinephrine; NIT, nitrite; NF-xB, nuclear factor kappa light chain

citrate synthase; DA, dopamine; DARPP-32, dopamine- and cyclic-AMP-regulated phosphoprotein of molecular weight 32000; ELT, escape latency; ERK, extracellular signal-regulated kinase; ETC,

electron transport chain; FST, forced swimming test; GABA, y-aminobuytric acid; GPx, glutathione peroxidase; GSH, reduced glutathione; GSSG, glutathione

S-transferase;

disulfide; GST, glutathione

morphological characteristics and viability of PC-12 cells were retained in cells treated with the date fruit extract. The finding suggests the ability of *Phoenix dactylifera* L. fruits $(1-1000 \ \mu g/mL)$ to protect neuronal cells against 3-NP-induced oxidative stress and biochemical changes under the HD *in vitro* model.

Psoralea corylifolia Linn. Psoralea corylifolia Linn, which belongs to Leguminosae family, is a well-known herbaceous legume that grows in China, Korea, and India. From ancient times, it has been used to ameliorate various ailments in both Chinese and Ayurvedic medicines. There are approximately a hundred bioactive compounds isolated from *Psoralea corylifolia* with flavonoid, meroterpene, and coumarin groups identified as notable bioactive phytochemicals having useful anti-inflammatory, radio-modulatory, antitumor, antiparkinsonian, and dopaminergic neuroprotective properties.³⁷

Previous *in vivo* studies have revealed that the seed extract of *Psoralea corylifolia*, furocoumarins, and psoralidin exert antidepressant effects.³⁸ In addition, it has been shown that *Psoralea corylifolia* seed extract (10, 50, and 100 μ g/mL) attenuated 3-NP triggered mitochondrial dysfunction in PC-12 neuronal cells in a concentration-dependent manner.³⁹ It restored the mitochondrial function by increasing mitochondrial respiratory capacities accompanied by an increased basal oxygen consumption rate.

Punica granatum Linn. Punica granatum Linn (Punicaceae) or pomegranate is one of the oldest edible fruits originating from the Mediterranean and Middle East areas and also growing in the parts of North Africa and Asia.⁴⁰ Traditionally, Punica granatum has been used to treat kidney disorders, urinary infection, thyroid dysfunction, atherosclerosis, and cardiovascular diseases.⁴¹ In Indian Unani medicine, Punica granatum is used to ameliorate diabetes mellitus. Due to its vast pharmacological activities, including anticancer, antioxidant, anti-inflammatory, antiulcer, and antilipoperoxidation, 40,42 the fruit has attracted enormous attention from researchers. A recent study reported that Punica granatum seed oil (40 μ M), which contains an abundance of unsaturated fatty acid acyl glycerols in the form of diynoic acid derivatives, can confer some protection against 3-NP-induced neurotoxicity in cultured PC-12 cells.43 The findings indicate suppression of lactase dehydrogenase activity and reduction of ROS, extracellular nitric oxide, and lipid peroxidation occurring in the PC-12 neuronal cells, which are attributed to the antioxidant activity of the seed oil as confirmed by in vitro radical scavenging assays. In addition, owing to the presence of the interrupted diynoic acid system by one methylene, the seed oil of Punica granatum exerted a superior antioxidant capacity to enhance antioxidant gene expression and neutralize ROS formation in PC-12 cells.⁴

In Vivo Studies. Calendula officinalis Linn. Calendula officinalis Linn, also known as marigold, belongs to the family of Asteraceae.⁴⁴ From ancient times, the flowers of Calendula officinalis have been applied as an herbal medicine for homeopathic purposes due to its ethnomedical value. Phytochemical screening indicates that Calendula officinalis flowers are rich in flavonoids (quercetin, isoquercetin, isorhamnetin, calendoflavoside, calendoflaside, calendoflavobioside, and rutin), carotenoids (α -carotene, β -carotene, and α -tocopherol), and quinones (phylloquinone and plastoquinone).⁴⁴

Shivasharan et al.⁴⁵ investigated the beneficial effects of *Calendula officinalis* extract in a HD *in vivo* model where constituents such as rutin and chlorogenic and ferulic acids

have been identified as the notable bioactive compounds that confer the neuroprotective actions. Interestingly, pretreatment with *Calendula officinalis* (100 and 200 mg/kg, po) markedly alleviated 3-NP-induced striatal oxidative damage, neuronal loss, and behavioral alterations. The observed protective propensity on the neurons is attributed to the antioxidant and anti-inflammatory potential of *Calendula officinalis* which can (1) normalize the endogenous antioxidant defensive enzymes and (2) reduce the oxidative and nitrative stress in the rat brain. Additionally, *Calendula officinalis* has some estrogenic properties, which may prevent the neuronal degeneration due to neurotoxicity since estrogen can increase antiapoptotic and decrease apoptotic gene expression, thus increasing the blood supply to brain regions and inhibiting the expression of pro-inflammatory cytokines.⁴⁵

Celastrus paniculatus Willd. Celastrus paniculatus Willd is an Ayurvedic medicinal plant from the Celastraceae family, also locally known as "Tree of life", Kangani, Malkangni, or Jyotishmati.⁴⁶ Accumulated reports have indicated that the extracted seed oil from Celastrus paniculatus possesses a series of neuroprotective activities together with having good potential as a memory enhancer. For instance, the study by Malik et al.⁴⁷ indicated that the seed oil from Celastrus paniculatus dose-dependently attenuated behavioral alterations and oxidative damage induced by 3-NP in the rat brain, which effects are largely attributed to its antioxidant activity that reduces both malondialdehyde (MDA) and nitrite levels. The levels of GSH, SOD, and catalase (CAT) activity are also restored in 3-NP intoxicated rats following treatment with Celastrus paniculatus (100 mg/kg, po). Additionally, a report by Godkar et al.⁴⁸ revealed that Celastrus paniculatus aqueous extract and seeds can protect neurons from glutamate-induced neurotoxicity by acting on NDMA receptors culminating in increased calcium (Ca^{2+}) influxes.

Centella asiatica (Linn) Urban. Centella asiatica (Linn) Urban, from the Umbelliferae family, is a native plant from India, Madagascar, Malaysia, Sri Lanka, and South Africa. For centuries, it has been widely used in Ayurvedic medicine to treat various ailments. The leaves of Centella asiatica contain triterpene saponins (sapogenins), madecassoside (madecassic acid), asiaticoside (asiatic acid), flavonols, and derivatives of caffeic acid, which are potent antioxidants. The various neuropharmacological potentials of Centella asiatica (5 mg/ kg, po) have been investigated in an HD in vivo model.^{49,50} In the study, prophylactic treatment using Centella asiatica extract conferred remarkable protection against 3-NP-induced protein and neuronal oxidative damage in all regions in the rat brain as evidenced by (1) attenuation of GSH, (2) total thiol (TSH) depletion, and (3) increased protein carbonyl levels. Additionally, Centella asiatica can prevent (1) membrane damage and (2) mitochondrial swelling and dysfunction, all of which contribute to the preservation of the metabolic rate in the mitochondria as well as maintenance of ETC integrity. Overall, these protective efficacies are ascribed to the ability of Centella asiatica to enhance the endogenous antioxidant status via the attenuation of oxidative stress and elevation of certain enzymatic and nonenzymatic antioxidant levels.

Convolvulus pluricaulis Choisy. In the past decades, Convolvulus pluricaulis Choisy. (Convolvulaceae), also known as Shankhpushpi, has been tested in the treatment of various central nervous system disorders.⁵¹ Convolvulus pluricaulis is reported to contain alkaloids, flavonoids (kaempferol derivatives), carbohydrates, volatile oil, phytosterol (β -sitosterol), scopoletin, 20-oxodotriacontanol, 29-oxodotriacontanol, and tetratriacontanoic acid used as an Ayurvedic nerve tonic called Medhya Rasayana.⁵² The tonic is believed to have antianxiety, antidepressant, anticonvulsant, memory enhancing, and sedative effects, useful in relieving insomnia, dyspepsia, fatigue or weakness, nervousness, and palpitation.⁵³

Various fractions of Convolvulus pluricaulis have been shown to confer some neuroprotective effects as confirmed in 3-NP intoxicated rat models. For example, Kaur et al.⁵⁴ indicated that pretreatment with methanolic extracts of Convolvulus pluricaulis (20 mg/kg, po) markedly ameliorated 3-NP-induced behavioral changes, which include the restoration of locomotor activity and rotarod and beam-walk performance. Additionally, the oxidative damage in a 3-NP treated rat model was reduced by attenuation of LPO levels and restoration of the defensive enzyme. In another study, Malik et al.⁵² reported that the standardized hydromethanol extract of Convolvulus pluricaulis (200 mg/kg, po), which contains scopoletin, significantly enhanced memory and cognitive function of 3-NP treated rats, decreasing the time latency (TL) in Morris water maze test (MWM). The findings suggest that the antioxidant activity of Convolvulus pluricaulis is exerted in the striatal region of the brain along with the simultaneous upregulation of protein synthesis in the hippocampus.

Ficus religiosa Linn. Ficus religiosa Linn (Moreceae), also known as peepal, pipal, or pimpala tree, is a large and deciduous tree with no or few aerial roots that grows in the tropical area throughout India.⁵⁵ It is also commonly cultivated in the vicinities of temples in South East Asia since the tree is religiously recognized as a sacred tree to both Buddhist and Hindu communities. The tree is traditionally used in a variety of the medical systems, including the Ayurveda, Homeopathy, Unani, and Siddha. In fact, the therapeutic potential of its various parts including the leaves, fruits, stem barks, aerial roots, seeds, vegetative buds, and latex has been confirmed.⁵⁶

The leaves of Ficus religiosa are of the major interest, because they contain lupeol, arginine, aspartic acid, tryptophan, proline, alanine, threonine, α -amyrin, campesterol, isofucosterol, stigmasterol, tyrosine, methionine, isoleucine, leucine, tannic acid, glycine, serine, hexacosanol, n-octacosane, n-nonacosane, *n*-hentricontanen, and valine contents.⁵⁷ These phytochemicals provide a range of pharmacological activities of Ficus religiosa, including antioxidant, antiacetylcholinesterase, antidiabetic, anti-inflammatory, analgesic, anticonvulsant, antimicrobial, antiamnestic, antiulcer, and proteolytic activities.^{57,58} Bhangale et al.⁵⁸ investigated the neuroprotective effect of Ficus religiosa at a high dose (400 mg/kg, po) against the 3-NP-induced HD model rats. Pretreatment with the ethanolic and ethyl acetate Ficus religiosa extracts significantly mitigated the behavioral, biochemical, and histological alterations purported to be contributed by its high antioxidant properties. The extracts decreased neuronal degeneration and neuroinflammation as well as ameliorated necrosis in the striatum. The tendency toward decreased oxidative stress in the rat brain along with the increased enzymatic and nonenzymatic antioxidant status indicates the neuroprotective properties of Ficus religiosa.

Ginseng Radix Rubra. Ginseng radix Rubra is a steamed root of *Panax ginseng*, which is also known as Korean red ginseng.⁵⁹ Among several types of *Panax ginseng*, Korean red ginseng has been reported to have useful antidiabetes, antihypertension, antinociception, and anticancer activities. Subsequently, investigation of extract of Korean red ginseng by Jang et al.⁶⁰ suggested that preadministration of the extract

(50, 100, and 250 mg/kg, po) dose-dependently alleviated neurologic impairment and lethality, accompanied by the decrease in striatal neuronal death and lesion area, followed by striatal degeneration. These protective effects may be contributed by the anti-inflammatory properties of the extract on striatal neurotoxicity. Another observation indicates that microglial activation, pro-inflammatory cytokine expression [interleukin-1 β (IL-1 β), interleukin-6 (IL-6), tumor necrosis factor α (TNF- α)], and elevation of iNOS are inhibited with pretreatment of Korean red ginseng extract, by deactivation of the mitogen-activated protein kinases (MAPK) phosphorylation and nuclear factor kappa light chain enhancer of activated B cells (NF- κ B) signaling pathways. Hence, ginseng radix warrants attention as a potential medicinal plant extract in HD *in vivo* model.

Luehea divaricata Mart. Luehea divaricata Mart (Tiliaceae) is a well-known natural product found in South America in the Southern regions in Brazil and also known as acoita-cavalo by the natives.⁶¹ Its leaves are reported to be rich in flavonoids, saponins, mucilage, and tannins, while the crude extract contains alkaloids, anthocyanidins, polysaccharides, carotenoids, and some fixed oils.⁶² For these reasons, Luehea divaricate has been claimed to have anti-inflammatory and antimicrobial activities and is primarily used to treat rheumatism, arthritis, blennorrhea, dysentery, leukorrhea, skin lesions, and any other gastrointestinal and respiratory infections.^{61,62} Courtes et al.⁶³ investigated the neuroprotective action of Luehea divaricata extract against HD-associated behavioral and biochemical changes as induced by 3-NP in the rat model. Interestingly, Luehea divaricata aqueous extract (500 and 1000 mg/kg, ig) tends to normalize the behavioral and motor deficits, improving the rotarod and locomotor performances. In addition, the biochemical alteration was attenuated corresponding to decreased oxidative stress and lipid peroxidation in the striatum and cortex, indicating the protective effect of this plant extract against HD in vivo model ameliorated via its strong antioxidant activity.

Panax quinquefolius Linn. Panax quinquefolius Linn, which is in the Araliaceae family, is also known as American ginseng. In the past decades, there have been many clinical studies worldwide supporting the benefits of Panax quinquefolius notably in neurodegenerative disease.⁶⁴ Enormous efforts have also been devoted to the investigation on ginsenosides, major active components of ginseng extract especially with regards to the stems and leaves, which have favorable pharmacological activities, including antioxidant, antineoplastic, and antistress properties.⁶⁵ For example, ginsenosides, isolated from ginseng extract, consist of a steroid-like four ring structure attached to sugar moieties⁶⁶ and are the major components with putative neuroprotective potencies.

Interestingly, American and Asian ginsengs contain unique relative amounts of ginsenoside groups, panaxadiols (Rb1, Rb2, Rb3, Rc, Rd, Rg3, Rh2, and Rh3) and panaxatriols (Re, Rf, Rg1, Rg2, and Rh1).⁶⁴ As reported by Lian et al.,⁶⁵ the isolated Rb1 (panaxadiol, 10 mg/kg, po) elicits putative neuroprotective activities, which included reduced striatal lesion volume and hippocampal neuron loss in 3-NP administered rats. In addition, Rb1 also ameliorates the behavioral changes and motor impairment in the HD *in vivo* model, suggesting its potent neuronal protection by virtue of its free radical scavenging properties.

Withania somnifera (Linn) Dunal. Withania somnifera (Linn) Dunal, from the Solanaceae family, is an important

Table 4. List of Natural Products (Isolated Compounds) with Potential against HD in Experimental Models

natural compounds	type of natural compounds	sources	refs
berberine	alkaloids	Coptis chinensis, Berberis species	Jiang et al. ³²
celastrol	triterpene	Tripterygium wilfordii	Cleren et al. ⁸⁰
dihydromyricetin	flavonoid	Ampelopsis grossedentata	Mu et al. ⁸⁵
embelin	para-benzoquinone	Embelia ribes	Dhadde et al. ⁸⁸
epigallocatechin gallate (ECGC)	polyphenol	Camellia sinensis	Kumar and Kumar ⁹⁰
esculetin		Foeniculum vulgare, Aesculus hippocastanum, Salvia officinalis	Karandikar and Thangarajan ⁹²
forskolin	diterpenoid	Coleus forskohlii	Mehan et al. ⁹⁴
genistein	isoflavone	soybeans	Menze et al. ⁹⁷
gintonin	lysophosphatidic acid	ginseng	Jang et al. ¹³⁹
lutein	carotenoid	green leafy vegetables, carrots	Binawade and Jagtap ¹⁰⁰
lycopene	carotenoid	tomatoes and tomato products	Sandhir et al. ¹⁷
naringin	flavonoid	citrus fruits	Gopinath and Sudhandiran ¹⁴¹
neferine	alkaloid	Nelumbo nucifera	Wong et al. ⁷⁴
nicotine		Nicotiana tabacum	Tariq et al. ¹⁰¹
onjisaponin B		Radix polygalae	Wu et al. ⁷⁷
praeruptorin C		Peucedanum praeruptorum	Wang et al. ¹⁰⁷
protopanaxatriol	ginsenosides	Panax ginseng	Gao et al. ¹⁴³
puerarin	isoflavonoid	Radix puerariae	Mahdy et al. ¹¹²
quercetin	flavonoid	fruits and vegetables	Chakraborty et al. ¹¹⁶
resveratrol	phytoalexin	grapes	Kumar et al. ¹¹⁸
S-allylcysteine	organosulfur compound	aged garlic	Elinos-Calderón et al. ¹²⁰
sesamol	phenolic compound	Sesamum indicum	Kumar et al. ¹²⁴
solanesol	polyisoprenoid alcohol	Nicotiana tabacum	Mehan et al. ¹²⁶
spermidine	polyamine	green vegetables, meat and milk products	Jamwal and Kumar ¹²⁸
sulforaphane	isothiocyanate	cruciferous vegetables	Jang and Cho ¹⁴⁵
T1-11		Gastrodia elata	Huang et al. ¹⁴⁷
tetramethylpyrazine	heterocyclic compound	Ligusticum wallichii	Danduga et al. ¹³⁰
<i>trans</i> - $(-)$ - ϵ -viniferin	stilbenoid	Vitis spp. (wild grape)	Fu et al. ⁷⁸
lpha-mangostin	xanthone	Garcinia mangostana	Pedraza-Chaverri et al. ⁷⁹
L-theanine	amino acid	Camellia sinensis	Thangarajan et al. ¹³⁴

medicinal plant. It has high value as an anabolic agent and is said to be health restorative, extensively applied in the Ayurvedic medicine system.⁶⁷ Withania somnifera is wellknown for its valuable therapeutic effects of anti-inflammatory, antimicrobial, antiarthritic, antistress, antidepressant, cardioprotective, and neuroprotective activities.⁶⁸ These biological activities are contributed by pharmacologically active phytochemicals, glycowithanolides, responsible for its protective effects against various diseases. Withania somnifera root extract (100 and 200 mg/kg, po) significantly improved motor coordination and increased grip strength of 3-NP treated rats in a dose-dependent manner.⁶⁹ The effect may be attributed to the restoration of the antioxidant enzyme level and the mitochondrial enzyme complex activity, indicating a possible antioxidant role of Withania somnifera against 3-NP-induced neurotoxicity in a HD in vivo model. Additionally, it has been postulated that Withania somnifera has the possible anabolic effect of reducing the 3-NP triggered ATP loss and energy impairment. It is plausible that the protective activity directly restores the cellular energetic balance and, in turn, prevents mitochondrial damage due to 3-NP-induced toxicity.

Zingiber officinale Roscoe. In the past 2000 years, Zingiber officinale Roscoe also known as ginger, has been utilized as a spice in food preparations around the world.⁷⁰ Traditionally, it has been applied as the main ingredient in Ayurvedic, Tibbi-Unani herbal, and Chinese medicine systems for treatment of asthma, rheumatism, gingivitis, hypertension, stroke, diabetes, and constipation.⁷¹ The ginger root and its aqueous extract possess certain polyphenol compounds, such as 6-gingerol and

its derivatives,⁷² suggesting the possible antioxidant properties of the plant to be a neuroprotectant against neurodegenerative disease. Sharma et al.⁷¹ confirmed the protective action of ginger root extracts in the 3-NP-induced HD animal model in which chronic treatment with ginger ethanolic extract (100 and 200 mg/kg, po) dose-dependently (1) improved memory, (2) mitigated motor and cognitive dysfunction, and (3) restored the biochemical changes with increased GSH and decreased nitrosative and oxidative stress levels in 3-NP treated rats. Taken together, the findings provide a new possibility of *Zingiber officinale*, which exerts its neuroprotective action via anticholinesterase potency.

Protective Effects of Natural Products (Isolated Compounds) against HD. The list of isolated natural compounds investigated as alternative medicines in HD preclinical models is outlined in Table 4, with their chemical structures depicted in Figure 5. Comprehensive information on their neuroprotective efficacy against HD from *in vitro* and *in vivo* studies are summarized in Tables 5 and 6, respectively.

In Vitro Studies. Neferine. Neferine is a bisbenzylisoquinoline alkaloid isolated from the seed embryo of *Nelumbo nucifera* Gaertn (Nelumbonaceae). *Nelumbo nucifera*, also known as lotus, is an edible and traditional Chinese medicinal plant that was initially cultivated in China.⁷³ Based on the report from Shenong from the Liang Dynasty, all of the plant parts (including the fruits, seeds, and roots) have a variety of medicinal effects and are commonly applied as antiaging and anxiety relief.³⁷ Due to its nontoxic nature, it is also widely used as a main ingredient in soup and tea preparations in



Figure 5. Chemical structure of naturally isolated compounds against HD in experimental models.

China. Neferine has potential as a neuroprotective agent against mHtt in PC-12 cells.⁷⁴ The data supports a working model for neferine (7.5 μ M) decreasing 3-NP-induced toxicity via induction of mammalian target of rapamycin (mTOR)–AMP-activated protein kinase (AMPK) [mTOR–AMPK] dependent autophagy by its activity on the autophagy related gene 7 (*Atg*7) with a higher expression of *Htt.*

Onjisaponin B. Onjisaponin B is one of the bioactive compounds that can be identified in radix polygalae (Yuan Zhi), the dried root of Polygala tenuifolia Willd, which is a popular Chinese medicinal plant traditionally used to promote tranquilization and mental alertness in China.⁷⁵ Radix polygalae extracts are reported to contribute to antipsychotic, antidepressant, memory, cognitive improvement, and sleeppromoting effects.⁷⁶ Therefore, to date, Radix polygalae extract is used to modulate neurodegenerative disorders and insomnia. Onjisaponin B was predominantly highlighted by Wu et al.⁷⁷ due to its neuroprotective potential in the HD in vitro model. Accordingly, Radix polygalae ethanolic extract markedly increased the conversion of LC3-I to LC3-II in PC-12 cells. Based on protein expression, the rate of LC3-II formation increased concomitantly with the existence of protease inhibitors, suggesting that onjisaponin B (25 and 50 μ M) inhibited neuronal damage via autophagy, which was activated due to the gene regulation of Atg7 via the mTOR-AMPK signaling pathway. The discovery successfully confirms that onjisaponin B has the potential to be a novel and effective autophagy enhancer without significant cytotoxicity, thus promoting the removal of mHtt and α -synuclein in PC-12 neuronal cells.

trans-(-)- ϵ -Viniferin. trans-(-)- ϵ -Viniferin is a unique collection of 22 types of stilbenic compounds composed of natural resveratrol monomers and oligomers. Fu et al.⁷⁸ reported that *trans*-(-)- ϵ -viniferin (EC₅₀ = 30 nM to 10 μ M) can protect cells from the detrimental effects of HD in an in vivo model. The neuroprotective effects are mainly exerted via the mediation of sirtuin-3 (SIRT3), a soluble protein responsible for controlling mitochondrial protein acetylation. In HD pathophysiology, mHtt induces the direct depletion of mitochondrial SIRT-3. In contrast, viniferin increases the levels of both SIRT-3 isoforms and activates the deacetylase activity of SIRT-3 in targeted substrate Mn-SOD, ultimately leading to enhancement of putative antioxidant activities. Subsequently, SIRT-3 deacetylates liver kinase B1 (LKB1) and directly induces AMPK activation, resulting in an increase of mitochondrial biogenesis that is accompanied by elevation of energy metabolism, which prevents the occurrence of mHtt induced mitochondrial dysfunction.78 Additionally, activated AMPK also (1) replenishes the levels of cellular nicotinamide adenine dinucleotide (NAD⁺) and (2) repetitively induces the activation of SIRT-3 and energy metabolism in order to protect the cells from oxidative damage.

α-Mangostin. α-Mangostin is a yellowish xanthone isolated compound from dried sap and bark of *Garcinia mangostana* (Guttiferae) is a tropical tree commonly cultivated in Malaysia, Indonesia, Thailand, Philippines, and Sri Lanka.⁷⁹ The pericarp of *Garcinia mangostana* has been widely used to treat diarrhea, abdominal pain, infectious wounds, chronic ulcers, and dysentery. α-Mangostin is claimed to have neuropharmacological activities against 3-NP-induced cultured cerebellar

а
tudies
S
Vitro
In
on
Based
Π
Ţ
agains
tential
Рс
with
(s)
punoduuc
Ŭ
(Isolated
Ś
Product
atural
Ž
of
Summary
s.
able
H

refs	Jiang et al. ³²	Jang et al. ¹³⁹	Kulasekaran and Ganapa- sam ¹⁸	Wong et al. ⁷⁴	Wu et al. ⁷⁷	Wu et al. ⁶⁶	Liu et al. ¹⁴⁴	Huang et al. ¹⁴⁶	Fu et al. ⁷⁸	Pedraza-Cha- verri et al. ⁷⁹	luctase; GSH, e; LPO, lipid (H); PC-12, pendent ^{<i>d</i>} Not
mode of action	induction of autopha- gy	inhibition of apopto- sis	Nrf2 activation	induction of mTOR– AMPK-dependent autophagy	clearance of mHtt in PC-12 cells via au- tophagy induction	inhibition of Ca ²⁺ sig- naling	activation of UPS and Atg pathways	induction of A _{2A} R signaling and PKA- dependent pathway	activation of AMPK via by increasing SIRT3	scavenge ROS	;; GR, glutathione rec actate dehydrogenas , NAD + hydrogen n. ^c Concentration de
molecular outcomes	Htt aggregation ↓, LC3-1/LC3-II ratio ↓	mHtt 4; neuronal death ↓	neurotoxicity J, LDH activity J; SOD †, CAT activity \uparrow , GPx †, GR †, GSH †; LPO †, ROS †; mitochondrial dysfunction J; activation of Nrf2 \uparrow	LC3-II †; mHtt 4, cell death 4	LC3-I to LC3-II conversion 1; Atg7 gene dependent; AMPK-mTOR inhibited	NMDA-induced Ca ²⁺ response ↓	GFPu 4, mHtt 4, neurotoxicity 4	mHtt aggregation 4, proteasome activity \uparrow	ROS \downarrow , NAD+/NADH ratio \uparrow , mitochondrial biogenesis \uparrow , SIRT3 and deacetylase activity \uparrow , activated AMPK \downarrow	$(^{1}O_{2})$ \downarrow , $O_{2}^{\bullet-}$ \downarrow , ONOO ⁻ \downarrow , cell damage \downarrow	GFP-UPS reporter; GPx, glutathione peroxidase 33-phosphatidylethanolamine conjugate; LDH, l D, nicotinamide adenine dinucleotide; NADH, proteasome system. ^b Most effective concentratio
model	genetically modified with Htt- 120Q	genetically modified with wild- type (STHIdh $^{Q/Q'}$) and mu- tant (STHIdh $^{QIII/QIT}$) cells	3-NP	genetically modified	genetically modified	glutamate	genetically modified with GFPu (GFP fused with degron CL1)	genetically modified	genetically modified with mHtt (N63-148Q)	3-NP	en fluorescent protein; GFPu, lic form of LC3; LC3-II, LC hroid 2-related factor 2; NA ide dismutase; UPS, ubiquitin
cell lines	HEK293 cells	STH1dh cells	PC-12 cells	cultured <i>Atg7</i> and <i>Atg7</i> deficient cells (WT-MEF); PC-12 cells	cultured <i>Atg7</i> and <i>Atg7</i> deficient cells (WT-MEF); PC-12 cells	cultured MSN	HeLa and HEK293 cells	PC-12 cells with mHtt	immortalized striatal precursor cells, Tet-Off PC-12 cells, N2a cells, cultured primary cortical neurons	cultured CGNs	ttophagy; CAT, catalase; GFP, gre N-terminal kinase; LC3-I, cytoso tro-2A; Nrf2, nuclear factor eryt ss; SIRT3, sirtuin-3; SOD, superox
EC_{50}	q	q	d	q	đ	đ	ą	~2.2 μM	30 nM to 10 μM	ą	icid; Atg, au NK, c-Jun ; N2a, neu sygen specie
IC_{50}	đ	а	q	12.8 μM	q	d	d	4.66 µM	а	5, 6, 8 μM	opropionic ; antingtin; J huntingtin , reactive ox
dose	5, 15, 50 ^b , 100 μM	0.1, 1, 10 $\mu \mathrm{g/mL}^c$	10 μM	7.5 μM	25, 50 μM	$0.01^{b}, 0.1^{b}, 1.0 \ \mu M$	2, 4, 8 μM ^c			$0-20 \ \mu M$	3-NP, 3-nitrc one; Htt, hu Htt, mutant ma-12; ROS
natural com- pounds	berberine	gintonin	naringin	neferine	onjisaponin B	protopanaxatriol	sulforaphane	T1-11	trans-(−)-€-vini- ferin	<i>α</i> -mangostin	^a Abbreviations: reduced glutathi peroxidation; ml pheochromocyto determined.

L

ACS Chemical Neuroscience

					outcomes			
e / duration e (days)		animal mod- els, sex	toxin	behavioral	biochemical	histopathological	mode of action	refs
		transgenic mouse: N171-82Q mice, male		motor coordination \uparrow , muscle strength \uparrow , body balance \uparrow	Htt aggregation 4; p62 ↓		induction of autophagy	Jiang et al. ³²
10		Lewis rats, male	3-NP, ip		Hsp70 expression 1, astrogliosis 4	striatal lesion volume 🔱	anti-inflammatory protec- tion by inducing Hsp70	Cleren et al. ⁸⁰
S		SD rats, male	3-NP, ip	body balance ↑, ELT ↓, TSTQ ↑	striatal glucose metabolism $\uparrow;$ MDA $\downarrow;$ SOD \uparrow	striatal injury ↓	antioxidant and antiapop- totic protection	Mu et al. ⁸⁵
; 14		Wistar rats	3-NP, ip	motor abnormalities J , locomotor count \uparrow , memory loss \downarrow , body balance \uparrow , hanging latency \uparrow	LPO J, GSH †, CAT activity †, GST †	brain lesions ↓	antioxidant defense sys- tem	Dhadde et al. ⁸⁸
), 14 po		Wistar rats, male	3-NP, ip	locomotor activity 1, grip strength 1	MDA 4, NIT 4, SOD 7, CAT activity 7; NADH, SDH, cytochrome oxidase activities 7; viable cells 7	striatal lesion volume 🕹	NOS inhibition	Kumar and Kumar ⁹⁰
0; 14		Wistar rats, male	3-NP, ip	mobility in OFT 1, TL 4, immobility times in FST 4, grip strength \uparrow	MDA 4, protein carbonyl 4; SOD 1, CAT activity 1, GPx 1; SDH activity 7, AChE 4		scavenging free radicals	Karandikar and Thangarajan ⁹²
, 15		Wistar rats, male	3-NP, ip	<pre>TL ↓, TSTQ ↑, locomotor count ↑, grip strength ↑, neurological score ↓</pre>	ATP \uparrow SDH activity \uparrow LDH activity \downarrow AChE \downarrow MDA \downarrow NIT \downarrow ; GSH \uparrow SOD \uparrow , CAT activity \uparrow	histological alteration by 3-NP (activation of AC via en- hancing cAMP/PKA/ CREB pathway	Mehan et al. ⁹⁴
∞		albino rats, female	3-NP, ip	locomotor activity 1, memory loss 4	ATP 1; LPO 1, GSH 1, CAT activity 1, AchE 1; iNOS 1, COX-2 4	brain histological features restored	antioxidant and antiapop- totic protection	Menze et al. ⁹⁷
- <i>?</i> *``		CS7BL/ 6NTac mice, male	3-NP, ip; AAV vector	neurological dysfunction \downarrow	SDH activity †; TNF-α, IL-1/ ² , IL-6, COX-2 and iNOS ↓; apoptosis ↓; activation of NF-xB ↓ microglial activation ↓	striatal cell death ↓	activation of LPA and Keapl–Nrf2–ARE pathway, inhibition of MAPKs and NF- <i>k</i> B	Jang et al. ¹³⁹
- - -	-	SD rats, fe- male	3-NP, ip	TL ↓, ELT ↓, locomotor activity ↑	NADH, SDH activities ↑, MTT ↑; GSH ↑, CAT activity ↑, LPO ↓; AchE ↓	mild focal gliosis	scavenging free radicals	Binawade and Jagtap ¹⁰⁰
1	NO.	Wistar rats, female	3-NP, ip	body balance ↑, TL ↓, memory loss ↓	NADH, SDH, cytochrome oxidase, F ₁ F ₀ synthase activities \uparrow ; NIT J, ROS J, LPO \downarrow ; SOD \uparrow ; TSH \uparrow , PSH \uparrow , NPSH \uparrow ; mitochondrial cytochrome $c \uparrow$, p53 expression \downarrow		↓ oxidative stress, ↓ mito- chondrial dysfunction	Sandhir et al. ¹⁷
-	4	Wistar rats, male	3-NP, ip	hind limb function \uparrow , grip strength \uparrow , print length \downarrow	mMP ↑, TIMP ↑; NF-xB ↓, GFAP ↓		modulation via expression of MMPs and GFAP	Gopinath and Sudhandiran ¹⁴¹
					GSH/GSSG ratio $\uparrow;$ activation of Nrf2 $\uparrow;$ TNF-a, COX-2 and iNOS \downarrow	histopathological alterations ↓	activation of Nrf2-medi- ated ARE gene pathway	Gopinath and Sudhandiran ^{14;}
					SOD f, CAT activity f, GPx f, GR f; GSH f, vitamin C and E f; ATPases f; Bax l, Bad l, Bcl-2 f; mitochondrial cytochrome c f, cytosolic caspase-3 activation \downarrow	histopathological abnor- malities in striatum ↓	antioxidant and antiapop- totic protection	Gopinath et al. ¹⁴⁰
-214		Wistar rats, female	3-NP, ip	locomotor activity †, grip strength †, body balance †, higher capacity angles in inclined plane test	DA †; GSH †; TH †	striatal lesions ↓	restoration of striatal DA	Tariq et al. ¹⁰¹
~	~	Wistar rats, male	3-NP, ip		LPO 4, protein carbonyl 4, SDH activity \uparrow		antioxidant defense sys- tem	Túnez et al. ¹⁰⁴

ACS Chemical Neuroscience

pubs.acs.org/chemneuro

Review

		refs	Wang et al. ¹⁰⁷	Gao et al. ¹⁴³	Mahdy et al. ¹¹²	Chakraborty et al. ¹¹⁶	Kumar et al. ¹¹⁸	Herrera-Mundo et al. ¹²¹	Kumar et al. ¹²³	Kumar et al. ¹²⁴	Mehan et al. ¹²⁶	Jamwal and Kumar ¹²⁸	Jang and Cho ¹⁴⁵	Liu et al. ¹⁴⁴	Danduga et al. ¹³⁰	Huang et al. ¹⁴⁷	Thangarajan et al. ¹³⁴
		mode of action	↑ expression of normal Htt, DARPP-32, and BDNF	↑ antioxidant defense system	antiapoptotic and anti-in- flammatory protection	scavenging free radicals, anti-inflammation pro- tection	antioxidant defense sys- tem, scavenging free radicals	↓ oxidative stress	scavenging free radicals	reverse mitochondrial en- zyme status	↓ mitochondrial dysfunc- tion	antioxidant and anti-in- flammatory protection	activation of Keap1– Nrf2–ARE pathway, in- hibition of MAPKs and NF-xB	activation of UPS and Atg pathways	↑ antioxidant, ↓ lipid peroxidation	activation of the adenosi- nergic system	antioxidant defense sys- tem
		histopathological	neuronal damage in the striatum J	normal morphology of nucleus, number of cells decreased in the striatum ↓		microglial proliferation ↓, astrocytes numbers ↑					rat brain optimally sized, cell nucleus and con- tinual cell membrane observable		striatal lesions 🗸		histological changes in striatum ↓, neuronal count in the hippocampus ↑		mild neuronal cell dam- age, reduced inflamma- tion, absence of ne- crosis and gliosis
outcomes		biochemical	DARPP-32 †, BDNF †; glutamate ↓	ROS [, SDH activity \uparrow , SOD \uparrow ; Hsp70 expression \uparrow ; Nrf2 activation \uparrow , HO1 and NQO1 expression \uparrow	cytosolic caspase-3 and cytochrome c $\downarrow,$ Bax $\downarrow,$ Bcl-2 $\uparrow,$ ATP $\uparrow,$ TNF- α $\downarrow,$ iNOS $\downarrow,$ NF-xB \downarrow	serotonin metabolism ↓, MAO-A ↓	GSH †; MDA ↓, NIT ↓; SDH activity ↑	LPO 4, SOD 7; MTT reduction ability \uparrow	ratio of GSH/GSSG $\uparrow,$ GST $\uparrow,$ LDH activity \downarrow	MDA J, NIT J, SOD f, CAT activity f, NADH, SDH, cytochrome oxidase activities f; viable cells f	ATP ↑, SDH activity ↑, GSH ↑, SOD ↑, CAT activity ↑; AChE ↓; MDA ↓, NIT ↓	LPO \downarrow , NIT \downarrow ; total NPSH \uparrow ; expression of TNF- α , IL-I β , IL-6 \downarrow ; catecholamine \downarrow ; GABA \downarrow , glutamate \downarrow ; purine \downarrow	SDH activity †; TNF-a, IL-1β, IL-6, COX-2 and iNOS ↓; apoptosis ↓; activation of JNK, ERK and NF-xB ↓ microglial activation ↓; Nrf2 ↑	GFPu 4, UPS function 1; Atg activity 1	SDH activity f, GSH f, SOD f, CAT activity f; GABA f, glutamate ↓; AChE ↓; MDA ↓	activation of $A_{2A}R$ f_{1} adenosine uptake J_{1} cAMP f_{1} striatal mHtt J_{1} proteasome activity \uparrow	LPO J, GSH 1; SOD 1, CAT activity 1; SDH activity 1
		behavioral	fall latency in rotarod 1, total distance in OFT 1, immo- bility times in TST and FST ↓	locomotor activity †		anxiety 4, motor coordination 1, gait abnormalities 4	TL ↓, VCM ↓, locomotor activity ↑		cognitive and memory performance 1	locomotor activity 1, muscle strength 1	ELT 1, TSTQ 1, grip strength 1, TL 1, locomotor count 1, balance beam-walking im- proved	locomotor activity and motor coordination ↑, TL ↓	neurological dysfunction \downarrow		TL \downarrow , spatial memory impairment \downarrow , locomotor count \uparrow , motor coordination \uparrow	deterioration of motor coordination ↓	TL J, grip strength ↑, mobility ↑
		toxin	3-NP, ip	3-NP, ip	3-NP, sc	3-NP, ip	3-NP, ip	3-NP, ip	3-NP, ip	3-NP, ip	3-NP, ip	3-NP, ip	3-NP, ip		3-NP, ip		3-NP, ip
	animal mod-	els, sex	CS7BL/6 mice	SD rats, male	albino Wistar rats, male	SD rats, male	Wistar rats, male	Wistar rats, male	Wistar rats, male	Wistar rats, male	Wistar rats, male	Wistar rats, male	CS7BL/6 mice, male	transgenic mouse with GFPu	Wistar rats, male	transgenic R6/2 mice, male	Wistar rats, male
	duration	(days)	×	S	5	4	12		14	14	15	21	œ		21		14
	dose (mg/ kg).	route	1.5, 3.0; po	5, 10 ⁶ , 20; po	200; ip	25, 50; ip	5, 10; ^c po	300; ip	5, 10, 20; po	5, 10, 20; po	5, 10, 15; ^c Po	5, 10; ^c po	2.5, 5.0 ⁶ ; ip		40, 80; ^c ip	0.05; ip	100, 200; [¢] Po
		natural compounds	praeruptorin C	protopanaxatriol	puerarin	quercetin	resveratrol	S-allylcysteine	sesamol		solanesol	spermidine	sulforaphane		tetramethylpyrazine	T1-11	L-theanine

Table 6. continued

nucleoporin 62; PSH, protein thiols; ROS, reactive oxygen species; SDH, succinate dehydrogenase; SOD, superoxide dismutase; TIMP, tissue inhibitor of

metalloproteinases; TNF-a, tumor necrosis factor a; TSH, total thiols; UPS, ubiquitin proteasome system; VCM, vacuous chewing movement. ^bMost effective dose. ^cDose dependent.

administration; p53, tumor protein 53; p62,

^aAbbreviations: 3-NP, 3-nitropropionic acid; AAV, adeno-associated virus; AcHE, acetylcholinesterase; ARE, antioxidant response element; Atg, autophagy; ATP, adenosine triphosphate; Bad, Bcl-2-

associated agonist of cell death; Bax, Bcl-2-associated X protein; Bcl-2, B-cell lymphoma 2; CAT, catalase; COX-2, cycloxygenase-2; ELT, escape latency; EKK, extracellular signal-regulated kinase; FST

forced swimming test; GABA, 7-aminobuytric acid; GFAP, glial fibrillary acidic protein; GFP, green fluorescent protein; GFPu, GFP-UPS reporter; GR, glutathione reductase; GSH, reduced glutathione; Htt, huntingtin; ip, intraperitoneal injection; IL-1 β , interleukin-1 β ; IL-6, interleukin 6; iNOS, inducible nitric oxide synthase; JNK, c-Jun N-terminal kinase; Keap1, Kelch-like ECH-associated protein 1; LPA, lysophosphatidic acid; LPO, lipid peroxidation; MAO-A, monoamine oxidase A; MDA, malondialdehyde; mMP, mitochondrial membrane potential; MMP, matrix metalloproteinase; NAD, nicotinamide adenine dinucleotide; NADH, NAD + hydrogen (H); NIT, nitrite; NPSH, non-protein thiols; Nrf2, nuclear factor erythroid 2-related factor 2; OFT, open field test; po, per os, oral

granule neurons (CGNs). The neuroprotective potential of α mangostin against 3-NP triggered neurotoxicity was closely linked to the attenuation of peroxynitrite anion (IC₅₀ = 5 μ M), superoxide anion (IC₅₀ = 6 μ M), and singlet oxygen (IC₅₀ = 8 μ M) generation in cultured CGNs,⁷⁹ thus suggesting that the scavenging of ROS is contributed by its antioxidant and antiinflammatory activities.

In Vivo Studies. Celastrol. Tripterygium wilfordii Hook (Celastraceae) is an ivy-like vine with a long story of use in Chinese traditional medicine to treat joint pain and fever. The root bark extract of Tripterygium wilfordii contains celastrol, a triterpene with known putative neuropharmacological effects against 3-NP-induced neurotoxicity in the HD animal model. In rats administered 3-NP, celastrol (3 mg/kg, ip) was demonstrated to suppress the production of pro-inflammatory cytokines and iNOS and microglial activation.⁸⁰ Additionally, it inhibits LPO in the inner and outer mitochondrial membrane by a direct free radical scavenging activity. Moreover, celastrol can induce the activation of DNA-binding activity of heat shock factor 1 (HSF1), followed by a higher expression of Hsp70 in the lateral brain striatum.⁸¹ Therefore, it was concluded that celastrol can reduce 3-NP-triggered astrogliosis markedly in the rat brain. Subsequently, it was confirmed that celastrol conferred complete protection against striatal lesions and neurotoxicity induced by 3-NP via its antioxidant and antiinflammatory abilities.

Dihydromyricetin. Dihydromyricetin is an isolated flavonoid abundantly found in Ampelopsis grossedentata, commonly known as vine tea (Tengcha).⁸² In Asia, vine tea has been traditionally used as a medicinal herbal tea and was believed to have health benefits in preventing and treating sore throat, common flu, jaundice hepatitis, and hypertension.⁸³ There are studies revealing the promising pharmacological properties of dihydromyricetin, including antioxidant, anti-inflammatory, hepatoprotective, anti-membrane lipid peroxidation, antithrombotic, anticarcinogenic, and antibacterial activities.^{82,84} The neuroprotective effects of dihydromyricetin have been demonstrated in a HD in vivo model involving 3-NP induced motor and cognitive deficits and striatal injury.⁸⁵ Dihydromyricetin (10 mg/kg, ip) improves energy metabolism in the striatum by decreasing MDA and increasing SOD level via the antioxidant defense system. Apart from this, dihydromyricetin has been shown to reverse 3-NP induced up-regulation of Bax and down-regulation of Bcl-2. Also, 3-NP induced cell apoptosis was markedly reduced in the dihydromyricetin treated groups. These findings indicate that dihydromyricetin offers mitigation of motor and cognitive impairments as well as striatal injury by virtue of antioxidant and antiapoptotic activities.

Embelin. Embelia ribes Burm belongs to the Myrsinaceae family. It is a natural product documented in Ayurvedic medicine from ancient times,⁸⁶ extensively used as a major ingredient in various Indian formulations. Embelin, chemically known as 2,5-dihydroxy-3-undecyl-1,4-benzoquinone, is the main active constituent from all the plant parts of Embelia ribes. It is a naturally occurring alkyl-substituted hydroxybenzoquinone. ⁷ Considerable evidence suggests that embelin possesses antioxidant, anticonvulsant, analgesic, anti-inflammatory, and antidiabetic properties.^{86,88} Additionally, emerging research on embelin has established its promising neuroprotective potential in a 3-NP-induced neurotoxicity rat model. Dhadde et al.88 revealed that the administration of embelin (10 and 20 mg/kg, po) can alleviate the damage to striatal neurons from 3-NP,

Table 6. continued

which in turn confers protection from the deleterious effects seen in behavioral and neurochemical alteration. The neuroprotective effect of embelin is attributed to (1) its free radical scavenging properties, (2) decreased lipid peroxide levels, and (3) increased nonenzymatic and enzymatic antioxidant levels. By normalizing the altered antioxidant defense system, embelin administration provides a possible mode of neuroprotection in animals with HD-like symptoms.

Epigallocatechin Gallate (ECGC). Camellia sinensis (Linn) is also known as green tea and is said to have originated from Southwest China. The young or mature leaves, stems, and shoots of *Camellia sinensis* are used to process different types of green tea products.⁸⁹ Owing to its attractive pharmacological effects, including anticarcinogenic, antimutagenic, antioxidant, antiproliferative, and neuroprotective properties,⁹⁰ green tea is the most popular herb used to make tea beverages all around the world. Green tea leaf extracts, L-theanine and epigallocatechin gallate (ECGC), have been confirmed to be effective in neuroprotection against 3-NP-induced toxicity of HD experimental model.

Epigallocatechin gallate (ECGC) is an important polyphenol present in green tea that can pass the blood-brain barrier (BBB). According to Kumar and Kumar,⁹⁰ the putative neuroprotective actions of ECGC (10, 20, and 40 mg/kg, po) involve attenuation of 3-NP-induced behavioral deficits and restoration of mitochondrial complex enzyme activities in order to preserve normal ATP levels. mHtt protein reduction and calcium influx inhibition are considered to be vital mechanisms in preventing neuronal damage. These protective actions can be explained by its antioxidant effects, which involve free radical scavenging, metal ion chelating, and ROS reduction. In addition, nitric oxide mediation may also contribute to the neuroprotective effects of ECGC by inhibiting nNOS and iNOS induction in the HD *in vivo* model.

Esculetin. Esculetin, a natural coumarin, is a secondary metabolite isolated from various plants, including Salvia officinalis, Aesculus hippocastanum, and Foeniculum vulgare.⁹¹ Esculetin has been reported to have anticancer, anti-ischemic, antidiabetic, and neuroprotective properties.⁹² Motivated by these pharmacological potentials of esculetin, its beneficial effects in the 3-NP-induced HD in vivo model have been investigated, where its most effective dose (25 mg/kg, po) significantly alleviated behavioral and biochemical alterations.⁹² In terms of behavioral improvement, the herb enhanced mobility in the open field test (OFT) and forced swimming test (FST), as well as the grip strength, of 3-NP treated rats. Additionally, lipid peroxidation was markedly reduced along with the restoration of antioxidant enzymes, suggesting that the possible neuroprotection occurs via antioxidant and free radical scavenging activity.

Forskolin. Coleus forskohlii, which belongs to the mint family, Lamiaceae, is an indigenous medicinal plant that originates from India and is locally known as mayani or makandi in Ayurveda.⁹³ Its tuberous root extract, forskolin, is the main constituent iterpenoid shown to possess the pharmacological effect on cardiovascular disease, hypertension, psoriasis, asthma, and eczema. Interestingly, intragastric administration of the natural product at 30 mg/kg markedly ameliorated the motor dysfunction and memory impairment of rats administered 3-NP.⁹⁴ It is plausible that attenuation of memory impairment mainly involved the brain cholinergic system, in which acetylcholinesterase (AChE) levels were reduced owing to the antioxidant capacity of forskolin in scavenging free radical, followed by an increase in ATP levels. Therefore, based on the above-mentioned observations, the vital role of adenyl cyclase (AC) activation in cAMP/CREB signaling pathway was investigated. Interestingly the results showed that AC/cAMP/PKA/CREB activation was responsible for attenuating HD-like symptoms in 3-NP treated rats.⁹⁵

Genistein. Genistein, one of the estrogenic compounds sharing structural features with 17β -estradiol (potent estrogenic compound), is a simple isoflavonoid occurring naturally in glycosylated forms.^{96,97} It is widely used as a functional and nutraceutical food due to its various therapeutic potentials like antihelminthic and antioxidant effects. Genistein can bind to estrogen receptors with up to 7–8 times higher binding affinity.⁹⁸

In recent years, high dietary intake of genistein has been linked to both memory and cognitive improvement in humans. These neuroprotective benefits are also demonstrated in an *in vivo* model associated with HD-like symptoms. Accordingly, systemic genistein administration (20 mg/kg, ip) increased retention latencies of 3-NP treated rats, further confirming the improvement in memory retention.⁹⁷ Furthermore, genistein protected neurons against oxidative stress-induced apoptosis by regulating the expression of Bax/Bcl-2. It is worth noting that genistein is effective in attenuating age-related memory and cognitive deficits via antiapoptotic and antioxidant activities.

Lutein. Lutein is a xanthophyll naturally found in plants such as marigold and green leafy vegetables and egg products.⁹⁹ When compared to other carotenoids, lutein warrants attention as a powerful antioxidant, a fact contributed by its unique chemical structure that possesses not only conjugated double bonds but also two hydroxyl groups on both ends, which strengthen its antioxidant activity.¹⁰⁰ Lutein (50 and 100 mg/kg, po) has been reported to have the ability to terminate the free radical reactions and protect neurons from 3-NP triggered oxidative stress in a rat model of HD-like symptoms in a dose-dependent manner.¹⁰⁰ The protective effects conferred restore the activity of the antioxidant defense enzymes and mitochondrial complex followed by the preservation of normal ATP function. Consequently, 3-NPinduced mitochondrial dysfunction is ameliorated. In addition, lutein normalizes the activity of AChE, thus improving memory function and cognitive task performance in 3-NP treated rats.

Lycopene. Another naturally occurring carotenoid, lycopene, is a red pigment present predominantly in tomatoes and tomato products. Lycopene is a potent antioxidant that can scavenge free radicals. Based on the findings of Sandhir et al.,¹⁷ the neuroprotective effects of lycopene against the HD *in vivo* model were ascribed to (1) neutralization of free radicals, (2) restoration of antioxidant status in the brain and (3) the modulation of lipid peroxidation in 3-NP treated rats. Lycopene (10 mg/kg, po) also normalizes ETC function, thereby alleviating mitochondrial dysfunction and causing a more controlled release of mitochondrial cytochrome *c* into the cytosol, which may activate apoptosis. Finally, it is suggested that the therapeutic effects of lycopene are also contributed by the suppression of activation of MAPKs and NF- κ B signaling pathways.

Nicotine. Nicotine, a natural alkaloid that originates from *Nicotiana tabacum* Linn (commonly known as tobacco), is composed of a pyrrolidine and pyridine ring. Moreover, nicotine crosses the blood-brain barrier and has some

neuroprotective effects as seen in 3-NP-induced HD *in vivo* models. Tariq et al.¹⁰¹ suggested that pretreatment of medium (0.5 mg/kg, ip) and high (1.0 mg/kg, ip) doses of nicotine significantly attenuated the depletion of striatal DA, followed by motor activity improvement in a dose-dependent manner. The pharmacological effects of nicotine likely to be mediated by the (1) involvement of neuronal nicotine acetylcholine receptor (nAChR) agonists¹⁰² and (2) stimulation of BDNF, nerve growth factor (NGF), glial cell line-derived neurotrophic factor (GDNF), as well as basic fibroblast growth factor (FGF-2).¹⁰³

Túnez et al.¹⁰⁴ revealed another neuroprotective mechanism of low dose nicotine (1.5 mg/kg, ip) in which it exhibited antioxidant effects by inhibiting oxidative damage induced by 3-NP, suggesting that nicotine sequesters ferrous ion (Fe²⁺) inhibiting the Fenton reaction. Moreover, nicotine enhances succinate dehydrogenase (SDH) activity by acting as a superoxide anion scavenger via inhibiting complex I of mitochondrial electron transport¹⁰⁵ and producing a monoaminergic neurotransmitter liberation on nACh receptors in cortical neurons. These protective actions synergistically restore energy metabolism in the rat's brain.

Praeruptorin C. Peucedanum praeruptorum Dunn is an herbaceous plant from Apiaceae family. Historically, the dried root of this plant, Peucedani Radix, is widely utilized in Chinese traditional medicine for the treatment of cough, airway infections, and hypertension.¹⁰⁶ Subsequently, it has been identified that praeruptorin C (PRA-C) is the main active constituent in dried root extract.¹⁰⁷ Research on PRA-C suggest that it exerts neuroprotective potency in attenuating the 3-NP-induced motor deficits, excitotoxicity, and depression in a HD *in vivo* model.^{107,108} In a separate study, Wang et al.¹⁰⁷ reported that the administration of PRA-C (1.5 and 3.0 mg/kg, po) effectively alleviated abnormal glutamate release while decreasing calcium influx. Additionally, PRA-C efficiently improves a series of behavioral and neurochemical alterations and histological damage in the striatum by preventing the loss of cellular viability from striatal neurons. Keeping this fact in view, the researchers conjectured the possible beneficial effects of PRA-C to be due to the upregulated expression of DARPP-32 (dopamine- and cyclic-AMP-regulated phosphoprotein of molecular weight 32000), Htt, and BDNF in the amygdala, which are mechanistically related to the regulation of emotion and motor function.¹⁰⁹

Puerarin. Radix puerariae (kudzu root), also known as Ge Gen in China, is the dried root of a medicinal plant, *Pueraria lobate* Ohwi. It is one of the earliest edible herbs in Oriental medicine.¹¹⁰ Puerarin (PUR) is a type of isoflavonoid [1.88–2.55% (w/w)] isolated from radix puerariae. PUR has been traditionally used for centuries in the treatment of cardiovascular diseases, hypertension, and diabetes mellitus.¹¹¹ Much evidence has confirmed the fact that PUR can penetrate the BBB immediately following intravenous administration thus conferring good neuroprotective potency.^{111,112} Owing to this fact, relevant studies involving PUR administration (200 mg/kg, ip) have been conducted in a 3-NP intoxicated HD *in vivo* model.^{112,113}

Accumulated results confirmed the neuroprotective effect of PUR on the brain tissue against 3-NP-induced neurotoxicity, in terms of reduced degree of hypothermia and brain injury with enhanced levels of antioxidative enzymes.¹¹³ The discovery is in agreement with previous reports in which PUR has been confirmed to protect hippocampal neurons from neurotoxicity

by (1) enhancement of CAT activity and GPx level, (2) reduction of LPO level, and (3) decreased formation of ROS. Taken together, this suggests that PUR provides neuroprotection by virtue of its antioxidant ability and neuromodulative effects. On the other hand, Mahdy et al.¹¹² reported that pretreatment with PUR prevented the increase in pro-apoptotic biomarkers by down-regulating the ratio of Bcl-2-associated X protein (Bax)/B-cell lymphoma 2 (Bcl-2) and attenuating caspase-3 activation thus conserving the antiapoptotic effects. In addition, treatment with PUR can (1) suppress NF- κ B activation, (2) inhibit iNOS expression, and (3) normalize ATP levels, ultimately restoring the energy for metabolism. Therefore, overall, the antiapoptotic and anti-inflammatory properties of PUR may protect the brain tissues from oxidative stress-induced inflammation and apoptosis.

Quercetin. Quercetin is a natural dietary flavonoid present abundantly in most edible fruits and vegetables including red onions, black or green tea, grapes, broccoli, blueberries, and cranberries.¹¹⁴ Quercetin possesses great pharmacological effects in preventing and treating cancer, cardiovascular disease and neurodegenerative disorders.¹¹⁵ Interestingly, quercetin can cross the BBB and exert its neuroprotective effect. Chakraborty et al.¹¹⁶ reported that quercetin (25 and 50 mg/kg, ip) can reduce 5-hydroxytryptamine (5-HT) levels markedly with a subsequent increase in striatal serotonin metabolism, possibly due to the inhibitory effect of monoamine oxidase A (MAO-A). Additionally, quercetin preserves mitochondrial function in 3-NP administered rats by a virtue of its antioxidant effects. Apart from this, the anti-inflammatory activity of quercetin has also been indicated by the decreased microglial activation and increased infiltration of astrocytes in the brain lesion core.

Resveratrol. Resveratrol, 3,4',5-trihydroxystilbene, is a phytoalexin naturally found in various plants such as peanuts, berries, and grapes.¹¹⁷ It is a nutraceutical that has useful anticancer, anticardiovascular disease, and antioxidant properties.¹¹⁸ Kumar et al.¹¹⁸ demonstrated that resveratrol (5 and 10 mg/kg, po) dose-dependently offers good protection against 3-NP-induced oxidative stress, improving both cognitive and motor performance of 3-NP treated rats. Due to the decreased oxidative stress, SDH activity was restored. Interestingly, resveratrol has been shown to decrease mitochondrial complex III activity in the respiratory chain of the rat's brain, which helps to directly maintain the stabilizing effect of the mitochondrial membrane, in turn, preserving mitochondrial function in the brain of 3-NP treated rats.

S-Allylcysteine. S-Allylcysteine is a compound that is an odorless, stable, and water-soluble organosulfur abundantly found in garlic.¹¹⁹ It is derived from cysteine, an amino acid in which an allyl group is attached to the sulfur atom.¹²⁰ The antioxidant effect of *S*-allylcysteine has been demonstrated in a 3-NP-induced toxicity *in vivo* model.¹²¹ *S*-Allylcysteine (300 mg/kg, ip) restores SOD activity and prevents lipid peroxidation by conferring primary protection of cells from 3-NP triggered oxidative stress. For the latter, the compound was found to prevent 3-NP-induced mitochondrial dysfunction and energy failure due to reduced ATP levels, thus suggesting that *S*-allylcysteine completely preserves cell survival and neuronal function from 3-NP-induced neuro- and excitotox-icities.

Sesamol. Sesamum indicum Linn (Pedaliaceae), also known as black sesame, is a herbaceous plant native to India and Africa. It is also cultivated in Asia mostly for its edible oil and seed. Traditionally, owing to its high degree of resistance to rancidity and oxidation, it is recognized as the "queen of oil seeds".¹²² Sesamol is one of the phenolic compounds that is mainly extracted from *Sesamum indicum*. To date, it has been commonly used as a popular dietary supplement worldwide due to its antioxidant value, which is effective in preventing hyperlipidemia, hypertension, and atherosclerosis and demonstrates antiaging and anticancer properties.³⁸

Motivated by the significant reported properties of sesamol, Kumar et al.^{123,124} have conducted investigations to explore the neuroprotective potency of sesamol against HD in the 3-NP-induced rat model. Pretreatment with sesamol (5, 10, and 20 mg/kg, po) conferred significant neuroprotection against 3-NP-induced motor and cognitive dysfunction and cellular alteration in different regions of the rat brain. Restoration of mitochondrial enzymes and nonenzymatic and enzymatic antioxidants has contributed to the amelioration of oxidative damage in the hippocampus, cortex, and striatum of the brain. The researchers hypothesized that the effect of sesamol in alleviating neuroinflammation is contributed by its antioxidant properties.

Solanesol. In fact, tobacco extracts and isolated compounds have good therapeutic potential against neurodegenerative disease owing to their anti-inflammatory, antioxidant, anesthetic, antispasmodic, sedative, and anticonvulsant properties.¹²⁵ In this direction, great interest has been directed toward several polyphenols present in tobacco for therapeutic purposes, including solanesol (SNL), a precursor of coenzyme Q10, which was shown to possess neuroprotective effects in the HD in vivo model based on 3-NP intoxication.¹²⁶ Interestingly, SNL (5, 10, and 15 mg/kg, po) dose-dependently increased ATP levels to a large extent thereby enhancing mitochondria function. By neutralizing free radical formation and increasing the enzymatic activity of coenzyme Q10, it restored the antioxidant activity and also attenuated the inflammatory damage in different areas in the rat's brain. Additionally, SNL also restored the cholinergic function and brain histopathological changes. Moreover, the findings of Mehan et al. support the neuroprotective potential of SNL in ameliorating 3-NP-induced memory and cognitive loss and motor incoordination.¹²⁶

Spermidine. Spermidine, one of the natural and ubiquitous polyamines, is an aliphatic polycation having a low molecular weight and nucleophilic centers. Meat, milk products, and green leafy vegetables are its major sources. Mounting evidence has revealed that spermidine has good antiaging effects, delaying the pathophysiology of neurodegenerative disor-ders.¹²⁷ Jamwal and Kumar¹²⁸ investigated the protective effects of spermidine in the 3-NP-induced in vivo model and found that pretreatment with spermidine (5 and 10 mg/kg, po) successfully ameliorated 3-NP triggered motor incoordination in a dose-dependent manner by (1) preventing the alteration of striatal neurotransmitters and (2) reducing the lipid peroxidation and oxidative stress followed by oxidative protein damage in rats. These protective effects could be wellcorrelated with the antioxidative potential of spermidine in scavenging free radicals in the rat brain. Owing to the antiinflammatory properties, spermidine is capable of deactivating the MAPK and NF-KB signaling pathways.^{127,128} Consequently, this phenomenon directly decreases the neuroinflammatory marker levels and striatal neuron degeneration in 3-NP treated rats, as well as prolonging the rat's lifespan.

Tetramethylpyrazine. Ligusticum wallichii Franchat (Chuan Xiong) is a traditional Chinese herb that belongs to the Umbelliferaceae family. It has been widely used in China in the treatment of cardiovascular and neurovascular diseases over the decades.¹²⁹ The active compound of Chuan Xiong, tetramethylpyrazine (TMP), has been reported to confer some neurotrophic and neurobiological effects while ameliorating HD-like symptoms in 3-NP-induced rats.¹³⁰ Therefore, attention is warranted for further investigation on TMP as a potential antioxidant against 3-NP neurotoxicity by inhibiting the elevation of lipid peroxidation (LPO) and scavenging the ROS.

In another study by Danduga et al.,¹³⁰ TMP (40 and 80 mg/ kg, ip) dose-dependently attenuated 3-NP excitotoxicity by increasing the γ -aminobutyric acid (GABA) levels as well as reducing glutamate levels. Moreover, it ameliorated memory impairment in rats by virtue of its antidepressant effect,¹³¹ attributed to the regulation of BDNF and the phosphorylation of CREB in the cAMP/PKA signaling pathway.¹³² Interestingly, the neuroprotective potency of TMP is attributed to its efficacy in restoring cholinergic functions, restoring striatal DA, and improving mitochondrial biogenesis in the brain, all of which contribute to the enhanced cognitive and motor performance.¹³⁰

L-*Theanine*. L-Theanine (γ -glutamylethylamide), which accounts for approximately 1.5% of the dry weight of tea leaves, is one of the major non-protein amino acids and is synthesized from glutamic acid and ethylamine in green tea leaves.¹³³ It is structurally similar to glutamate and GABA. L-Theanine can cross the BBB via a larger neutral amino acid transport system. It diminished the mitochondrial dysfunction in rats systemically administered 3-NP by preventing the reduction of LPO level and SDH activities, as well as by restoring the levels of enzymatic antioxidants, as confirmed by Thangarajan et al.¹³⁴ On the other hand, subchronic treatment with L-theanine (100 and 200 mg/kg, po) dose-dependently enhanced GABA synthesis, which triggers relaxation. Collectively, L-theanine mitigates 3-NP-induced oxidative stress by inhibiting NF- κ B activation, which is accompanied by upregulated expression of BDNF.¹³⁵ Finally, it can be concluded that L-theanine offers its neuronal protection against 3-NP triggered neurotoxicity in rats via anti-inflammatory and antioxidant activities.

In Vitro and In Vivo Studies. Berberine. Berberine is a protoberberine alkaloid that can be derived from the bark and roots of Berberis sp. or Coptis chinenses. For over six decades, berberine has been widely used in Chinese medicine to treat diarrhea due to bacterial infection. To date, it has been indicated in the treatment of cardiovascular disease, hypercholesterolemia, inflammation, and diabetes. Its benefits include (1) a high tolerance oral dose [50% lethal dose $(LD_{50}) > 5 \text{ g/kg}$ and (2) the ability to cross BBB. Due to these reasons, there is high interest to investigate its neuroprotective potential in both *in vitro* and *in vivo* models.³² The findings indicated that berberine (40 mg/kg, po) can improve the motor function of transgenic mice in a HD in vivo model. It also confers neuroprotection by activating Nrf2 and glucagon-like protein (GLP-1) and inhibiting MAO-B and AChE. Interestingly, berberine (50 μ M) can also enhance the autophagy process in parallel with increased degradation of mHtt in a HD in vitro model.

Gintonin. Gintonin is a novel lysophosphatidic acid (LPA)-ginseng protein complex naturally derived from

ginseng.¹³⁶ It contains an exogenous G-protein-coupled lysophosphatidic acid receptor (LPAR) ligand capable of repressing inflammation. Studies have reported that oral administration of gintonin exhibited antidementia and antimetastatic effects¹³⁷ and enhanced memory, learning, and physical stamina.^{138,139} However, the anti-inflammatory potential of gintonin remains unclear. Motivated by this reason, Jang et al.¹³⁹ investigated the anti-inflammation activity of gintonin against striatal toxicity in both in vitro and in vivo HD models. The findings from the HD in vivo model reveal that preadministration, coadministration, and onset administration of gintonin (100 mg/kg, po) reduce striatal cell death and attenuate neurological impairment by mitigating 3-NPinduced mitochondrial dysfunction, expression of pro-inflammatory cytokines, iNOS and cycloxygenase-2 (COX-2), and microglial activation. It was suggested that gintonin exerts its protective mechanism via the (1) inhibition of MAPKs, (2) NF- κ B signaling pathways, (3) activation of LPAs, and (4) Nrf-2 pathways.¹³⁹ Additionally, the beneficial effects of gintonin (0.1, 1.0, and 10.0 μ M) in a concentration-dependent manner were also reported in the adeno-associated virus (AAV) vector-infected STHdh cell HD in vitro model, reducing the neurological impairment via the mechanism of reduced formation and expression of mHtt aggregates.

Naringin. Naringin, 4,5,7-trihydroxy-flavonone-7-rhamnoglucoside, is an important major dietary flavanone glycoside found in citrus and grapefruits including Citrus unshiu, Citrus sinensis, Citrus paradise, Poncirus sp., and Artemisia selengensis due to its purported antioxidant, anti-inflammatory, antiatherogenic, and anticarcinogenic effects.¹⁸ The neuroprotective potential of naringin against HD-mediated neurotoxicity has been shown in various studies.^{18,140–142} The discoveries from both in vitro and in vivo reports support the hypothesis that naringin is the most potent Nrf2 inducer and protects neuronal cells against 3-NP-induced neurotoxicity by activating Nrf2.¹⁸ Collectively, naringin (80 mg/kg, po) increases the nuclear accumulation of Nrf2, which directly induces Nrf2regulated ARE gene expression. Subsequently, the activation of Nrf2 exerts a series of protective effects in multiple pathways including induction of the expression of phase II antioxidant genes, glutathione S-transferase P1 (GSTP1), γ -glutamylcysteine synthetase (GCS) mRNA, NQO1, and HO1.

On the other hand, naringin (10 μ M) decreases the expression of 3-NP-induced matrix metalloproteinase-2 (MMP2), matrix metalloproteinase-9 (MMP9), NF-kB, and glial fibrillary acidic protein (GFAP), which indicates its potent neuroprotective effects in preventing striatal toxicity and oxidative damage in 3-NP treated rat brain as well as PC-12 cells.¹⁴¹ In addition, naringin elicits its antioxidant effects by (1) potentiating the endogenous antioxidant defense capacity through an increase of SOD and GPx levels and CAT activity in cells and (2) preserving the mitochondrial respiratory chain integrity via a decrease in mitochondrial complex enzyme activities. Apart from the antioxidant properties, naringin possess antiapoptotic effects that can protect cells from neuronal apoptosis by decreasing the expression of Bcl-2associated agonist of cell death (Bad) and Bax concomitant with a decrease in caspase-3 and cytochrome c.¹⁴⁰ Taken together, these protective activities contribute to cellular protection against 3-NP-induced hazardous effects.

Protopanaxatriol. Panax ginseng C.A. Mayer, ginseng from the Araliaceae family, is an ancient herb widely used in traditional Chinese medicine. Ginsenosides are important components responsible for the neuroprotective actions of ginseng, as also mentioned before. There are more than 30 different kinds of ginsenosides that can be isolated from *Panax ginseng* and are classified based on their chemical structures into (1) protopanaxatriols (Rg1, Rg2, and Re) and 2) protopanaxadiols (Rb1, Rb2, Rc, Rd, and Rg3).

Protopanaxadiol Rb1 can pass through the BBB. In an *in vitro* HD assay, Rb1, Rc, and Rg5 conferred some protection to cultured MSNs against glutamate-induced neurotoxicity.⁶⁶ The findings revealed that Rb1, Rc, and Rg5 at low concentrations (0.01 and 0.01 μ M) successfully attenuated glutamate-induced apoptosis that accompanied the inhibition of neuronal Ca²⁺ signaling. Therefore, it is plausible that the neuroprotective effects of protopanaxadiols on the glutamate-induced HD *in vitro* model occurs due to the inhibition of the Ca²⁺ signaling pathway.

On the other hand, Gao et al.¹⁴³ conducted an *in vivo* study investigating protopanaxatriol (PPT) (10 mg/kg, po), which was found to confer some neuroprotection against 3-NPinduced behavioral, biochemical, and histological changes in Sprague-Dawley rats. PPT elicits its antioxidant effects via a series of mechanistic actions, including (1) free radical scavenging, (2) reduction of overproduced ROS, (3) inhibition of neuronal oxidative stress-induced reactions, and (4) elevation of activated endogenous antioxidant enzymes. Through these mechanisms, PPT directly restores the activity of the mitochondrial complex enzymes, subsequently protecting the mitochondrial function. By acting as a potent antioxidant agent, PPT upregulates the entry of nuclear factor erythroid 2-related factor 2 (Nrf2) into the nucleus along with the expression of heme oxygenase 1 (HO-1) and NAD(P)-H:quinone oxidoreductase 1 (NQO-1) which ultimately protects the neurons from 3-NP-induced neurotoxicity or striatum damage.

Sulforaphane. Sulforaphane, 1-isothiocyanato-4-(methylsulfonyl)-butane, is a dietary isothiocyanate correlated with antioxidant, antiapoptosis, antigenotoxicity, anticancer, and antimicroglial activation effects. It is also known as a chemopreventive agent most abundantly found in various cruciferous vegetables including cauliflower, brussels sprouts and broccoli. A mechanism of action of sulforaphane has been proposed by Liu et al.¹⁴⁴ based on an *in vitro* study. The study found that sulforaphane (2, 4, and 8 μ M) concentrationdependently activates Atg pathways concomitant with autophagy activity, therefore synergistically promoting the removal and degradation of unwanted substrates in bulk.¹⁴⁴ Since sulforaphane can cross the BBB, it has dual effects of (1)maintaining protein homeostasis and (2) suppressing free radicals to protect the neuronal cells against the toxicity of 3-NP.

The compound is also reported to mitigate 3-NP-induced striatal neurotoxicity in a HD *in vivo* model.¹⁴⁵ In the study, pretreatment with sulforaphane (5 mg/kg, ip) effectively ameliorated the neurological impairment along with striatal lesion volume triggered by 3-NP intoxication by the activation of Kelch-like ECH-associated protein 1 (Keap1)–Nrf2– antioxidant response element (ARE) [Keap1–Nrf2–ARE] pathway and the inhibition of MAPK and NF-*x*B pathways. Together with these signaling pathways, sulforaphane suppresses 3-NP-induced oxidative stress, SDH activity, microglial activation, apoptosis, the elevation of iNOS, expression of COX-2, and pro-inflammatory cytokines (IL-1 β , IL-6, and TNF- α). Additionally, the activated Nrf2–ARE

pubs.acs.org/chemneuro



Figure 6. Possible neuroprotective mechanism of natural products against HD in experimental models.

pathway sequentially induces phase II metabolizing antioxidant enzymes to exert antioxidant effects. The degradation of mHtt and ubiquitinated proteins via the UPS pathway is also enhanced by the peripheral injection of sulforaphane.¹⁴⁵

T1-11. Gastrodia elata Blume is from the Orchidaceae family. It has a long history recorded in Chinese Pharmacopeia for 1500 years. The dried tuber of *Gastrodia elata* is used to treat dizziness, headaches, limb numbness, spasms, and convulsive illness such as tetanus and epilepsy.^{146,147} To date, the literature reports two compounds, especially vanillin and gastrodin, that possess anticonvulsive, antiepileptic, and sedative properties. Two other active compounds, namely, bis(4-hydroxylbenzyl)sulfide and *N*-(4-hydroxybenzyl)adenine riboside (T1-11), have also been shown to protect PC-12 cells from apoptosis, confirming the central protective effects of *Gastrodia elata*.

Huang et al.¹⁴⁷ demonstrated the neuroprotective effects of T1-11 (4.66 μ M) by an *in vitro* assay, showing decreased mHtt aggregation and increased proteasome activity in neuronal PC-12 cells indicating its potential in HD. Similar neuropharmacological effects of *Gastrodia elata* extract were also reported in a transgenic HD *in vivo* model. T1-11 (0.05 mg/kg, ip) acted via the adenosinergic system in synapses by activating the adenosine A_{2A} receptor (A_{2A}R) and is therefore recognized as a potent adenosine analog that performs dual functions of (1) activating A_{2A}R and (2) blocking the adenosine transporter. Both activities synergistically inhibit endogenous adenosine uptake to exert a neuroprotective mechanism. Additionally, activation of A_{2A}R also effectively facilitates the function of certain neurotrophic factors, including BDNF, GDNF, and FGF.¹⁴⁷

POTENTIAL NEUROPROTECTIVE MECHANISMS

To gain insight into the underlying mechanism of natural products and compounds on HD-like symptoms (Figure 6), there were numerous *in vitro* and *in vivo* studies conducted.

They generally involved the antioxidant defense system, scavenging free radicals, neutralization of ROS, reduction of oxidative stress, preservation of mitochondrial function, antiinflammatory protection, inhibition of apoptosis, and induction of autophagy. ROS, including hydroxyl radicals (OH[•]), superoxide (O_2^-), and hydrogen peroxide (H_2O_2) are continuously generated during cellular aerobic respiration.^{148,149} ROS (1) plays a decisive role in modulating the biological processes of nerve cells in counteracting endogenous antioxidant defense status in the CNS and OH[•] involving multiple biological reactions and (2) is responsible for the oxidative damage to proteins, DNA, and lipids, while O_2^- take part in the production of H_2O_2 .

Physiologically, the levels of ROS generated are in equilibrium with the cellular antioxidant status. Nevertheless, when the level overwhelms the endogenous antioxidant capacity, a state of oxidative stress, cellular oxidative damage in the brain might ensue.¹⁴⁸ This condition is one of the convergent mechanisms closely correlated with the pathogenesis of HD and other neurodegenerative diseases since the CNS region is rich in polyunsaturated fatty acids where a high consumption of oxygen leads to a higher vulnerability to oxidative stress.

Interestingly, natural products (*Punica granatum* and tetramethylpyrazine) have been demonstrated to possess antioxidative properties in 3-NP treated brain and in PC-12 cells.^{43,130} Mounting evidence has indicated that these natural products are capable of sequestering free radicals, reversing the decreased level of enzymatic and nonenzymatic antioxidants, and in turn attenuating the elevated level of cellular oxidative stress. In reacting to the oxidative stress, a compensatory mechanism of endogenous antioxidative defense system is also induced by activation of Nrf2. The activated Nrf2–ARE signaling pathway sequentially induces the metabolizing antioxidant enzymes in exerting antioxidant effects by upregulating the enzymatic antioxidants (SOD, CAT, and

GPx) and nonenzymatic antioxidants (GSH), to neutralize the excess ROS in the brain. Many studies revealed that SOD and its isoforms, Zn-, Fe-, Cu-, and Mn-dependent SOD (Zn-SOD, Fe-SOD, Cu-SOD and Mn-SOD), preferably detoxify O_2^- by transforming it to H₂O₂. To detoxify H₂O₂, CAT is responsible for converting it to H₂O. In addition, GSH, GPx, γ -glutamyl-cysteinyl-ligase (GCL), glutathione reductase (GSSR), and glutathione transferase (GST) also play a vital role in detoxifying H₂O₂ to H₂O.¹⁵⁰ Collectively, lipid peroxidation is reduced and these activities can directly protect the distinctive cells and neurons against 3-NP-induced oxidative damage.⁴⁷ Furthermore, AChE level is reduced owing to the antioxidant capacity of forskolin, lutein, and berberine, suggesting the anticholinergic ability of these natural products in offering neuroprotection.^{32,100}

Mitochondria are the main engine for ATP production, which involves a series of oxidative phosphorylation reactions in the mitochondrial respiratory chain. High oxygen and glucose consumption in the brain make neurons more dependent on ATP generation within the mitochondria. Along the mitochondrial respiratory chain, mitochondrial complex enzymes I, II, IV, and V play a pivotal role in ATP synthesis. Interestingly, some natural products like Withania somnifera, epigallocatechin gallate, and sesamol have been shown to upregulate the enzyme activity of NADH, SDH, cytochrome oxidase, and F_1F_0 synthase.^{69,124,134} The restored mitochondrial complex enzyme activities are capable of normalizing ATP synthesis. Moreover, certain natural compounds such as genistein and naringin can mitigate glutamateinduced Ca2+ overloads on the NDMA receptor, thereby preserving ATP production.^{48,66} Additionally, natural products like Psoralea corylifolia can restore mitochondrial membrane potential $(\Delta \Psi)$ by normalizing Ca²⁺ influxes and ATP synthesis, thus preventing inner and outer membrane damage and mitochondrial dysfunction, as well as preserving the metabolic rate in mitochondria along with maintaining ETC integrity.39

It is noteworthy that a number of natural products like Calendula officinalis Linn and puerarin possess anti-inflammatory activities that are responsible for decreasing neuroinflammation.^{45,112} The pathological mechanism of HD extensively involves the elevation of pro-inflammatory markers (TNF- α , IL-1 β , and IL-6), which directly activate the NF- κ B signaling pathway together with the activation of microglia and astrocytes. In response to the inflammatory conditions, certain natural products can impact the microglia by inhibiting the activation of glial cells and astrocytes and by blocking the NF- κB pathway, thereby attenuating the delivery of proinflammatory cytokines to the immune system in HD preclinical models.^{80,145} Apart from inhibiting inflammatory pathways, the reduced expression of these cytokines, especially TNF- α can inhibit HD-induced apoptotic pathways as a result of elevated levels of TNF- α and its binding on TNFr1-type receptors, leading to the activation of apoptotic cascades.

A plethora of studies suggested that natural products such as puerarin elicit antiapoptotic effects by (1) upregulating the expression of Bcl2 and (2) decreasing the expression of Bad and Bax, both concomitant with the blocking of cytochrome *c* release via the deactivation of caspase-3.^{18,112} BDNF, NGF, GDNF, and FGF-2 are a group of neurotrophins that are beneficial for neuronal survival by preventing apoptosis. In this context, natural products have a role in stimulating the expression of these neurotrophins, DARPP-32, and Htt, thus

mechanistically inhibiting the neuronal apoptosis.^{103,146} Many natural products have an antiapoptotic ability, which may also contribute to attenuation of decreased levels of catecholamines (dopamine, norepinephrine, and serotonin) in striatum nuclei.¹⁵¹

Autophagy is a cellular self-degradation and removal process for dysfunctional cytoplasmic components. It was reported that natural compounds (onjisaponin B, neferine, and sulforaphane) have been shown to reduce 3-NP-induced neurotoxicity via the induction of mTOR-AMPK dependent autophagy activities.^{74,77,145} The findings support a working model of natural products to ameliorate the neuronal damage by activating the autophagy activities dependently on the gene regulation of Atg7 via the mTOR-AMPK signaling pathway. Consequently, the aggregation of mHtt was shown to be reduced by autophagy in cultured PC-12 cells, and the cognitive function of rats in the HD in vivo model was enhanced. Accumulating literature provides proof of neuroprotective potential from various natural products to attenuate HD-like symptoms under in vitro and in vivo models via single or combined neuropharmacological mechanisms.

CHALLENGES AND OPPORTUNITIES IN THE FIELD OF NATURAL PRODUCTS AGAINST HD

Medicinal plants and their isolated compounds have potential to be a valuable resource for drug discovery against HD. In this review, although several preclinical experiments have shown therapeutic promise, human HD clinical trials remain a challenge and are severely lacking. In translating the promising preclinical research to clinical applications, there are several difficulties and constraints such as poor water solubility of the natural products, their physicochemical properties, prompt metabolism, and low bioavailability. In addition, the existence of the BBB restricts their passage to the brain and some specific sites of action.

Nevertheless, since some reported isolated compounds in this review (lutein, lycopene, naringin, quercetin, resveratrol, Sallylcysteine, and sulforaphane) are widely found in fruits and vegetables commonly consumed, phase I clinical trials can be intiated, since these food products are deemed as safe for human consumption. Additionally, most of the medicinal plants reviewed have been traditionally used for many years in Ayurvedic medicine, traditional Chinese medicine, and other traditional medical systems. Another recent trend is the enrichment of foods and beverages, where fortification techniques using natural products may be a potential prevention strategy for HD. Although natural products should be tested in preclinical studies before evaluating their effect on humans in randomized clinical trials, which can help assess safety, tolerance, and efficient therapeutic doses for disease treatment, this poses a challenge.

Low bioavailability is one of the key drawbacks associated with natural products, which restricts their delivery to the target for pharmacological action. The use of nanotechnology and nanocarrier-based methods in the delivery of natural products may help solve these problems and enhance therapeutic responses while improving their efficacies. In fact, the integration of nanoparticles into brain-focused drug delivery systems (DDSs) is a promising strategy to increase the bioavailability and transport of natural products through the BBB.^{152–156} DDSs can protect natural products from biological degradation when the molecules are transported into the brain.¹⁵⁷ Thus, low doses of natural products can steadily

be released into the brain for enhanced efficacies. Nose to brain drug delivery (NBDD) is another useful method for improving the absorption, bioavailability, and therapeutic effects of natural products for treating brain disorders.¹⁵⁸ Growing evidence has indicated that nasal administration is a possible direct route to prevent BBB from blocking transport of drugs into the brain and has high bioavailability for amelioration of brain diseases.¹⁵⁸ In addition to the above-mentioned DDSs, new pharmaceutical technologies such as liposomal nanoencapsulation, polymeric micelles, cyclodextrins, nanosuspensions, and nanoemulsions are needed to increase the bioavailability of natural products, resistance to metabolic processes, and passage through the BBB.^{159–164}

In another aspect, drug metabolism and pharmacokinetics (DMPK) research is essential for understanding the efficacy and safety of natural products against HD. The reported compounds may be structurally modified to improve the DMPK properties, and such modifications may help to increase the activity strength and selectivity, improving solubility and partition coefficient, increasing metabolic and chemical stability, modulating pharmacokinetic parameters, and removing or alleviating toxicity and adverse reactions. To accomplish these multidimensional operations, sophisticated syntheses and skillful preparation of complicated molecules are essential. Finally, the mutual cooperation and interaction between organic and medicinal chemists are crucial in modifying natural products for commercial use through drug research and development. For the drug discovery process of HD, the isolated compounds listed in this review must be filtered using the Lipinski and Veber rules on the basis of druglike properties.^{165,166} Further, it is important to filter the listed compounds by removing substructures with Pan Assay Interference Compounds (PAINS) to avoid false positives.^{167,168} All these findings together can be used to discover a potential lead compound against HD based on common natural products.

METHODS

For this review, relevant studies were collected from several scientific databases including PubMed, ScienceDirect, Scopus, and Google Scholar. The categories of keywords used for the search included

- (a) "Natural product" OR "Medicinal plants" OR "Plant extract" AND "Huntington's disease".
- (b) "Natural compounds" OR "Phytochemicals" AND "Huntington's disease".

After screening literature from 2005 to the present, a total of 14 plant species and 30 isolated natural compounds that have been investigated against HD based on either *in vitro* or *in vivo* models were included in the present review.

CONCLUSION AND FUTURE PERSPECTIVES

This review presents a comprehensive account of the neuroprotective efficacy of various natural products against HD experimental models. Fourteen medicinal plants and 30 isolated compounds described herein have been shown to be effective against HD. Most of the medicinal plants listed in this review are traditional medicinal plants used for the treatment of brain disorders. The majority of the isolated compounds mentioned in this review are primarily from common plant species, are present or incorporated into traditional medicinal plants, or are found in food sources including in fruits, herbs, and spices. Since HD is a multifactorial disease, only natural products identified with distinct therapeutic mechanisms were included in this review.

Interestingly, promising in vitro and in vivo evidence has accumulated that Anemarrhenae asphodeloides (0.5 and 1.0 μ g/ mL) and Centella asiatica (5 mg/kg, po) are potentially effective at low concentrations or doses compared to other medicinal plants to protect against 3-NP-induced HD. Among the isolated compounds mentioned, proropanaxatriol (0.01 and 0.1 μ M) and nicotine (0.25, 0.50, and 1.0 mg/kg, ip) are potentially effective when used in low concentrations or doses against 3-NP-induced HD. Apart from these, all the other natural products reported in this review also can ameliorate altered behavioral, biochemical, and histopathological parameters, indicating their potential effectiveness based on in vitro and in vivo HD models. For most of the reported natural products, the possible modes of protection have also been extensively studied. Conclusively, accumulated evidence has implicated the model of natural products in HD preclinical models working mainly through the antioxidant defense system, preservation of mitochondrial function, anti-inflammatory protection, inhibition of apoptosis, and induction of autophagy. Nevertheless, several perspectives on the application of natural products for the prevention and treatment of neurogenerative potential are suggested. In addition to conducting further research to understand how natural products exert their therapeutic effects on HD, it is also necessary to undertake additional experimentation to target only certain natural products acting on the brain. Although natural products have vast therapeutic potential against HD, most are confirmed only in early phases of study. Randomized controlled trials are required to further strengthen the claims.

AUTHOR INFORMATION

Corresponding Author

Mahendran Sekar – Department of Pharmaceutical Chemistry, Faculty of Pharmacy and Health Sciences, Universiti Kuala Lumpur Royal College of Medicine Perak, Ipoh 30450, Perak, Malaysia; orcid.org/0000-0002-3022-6137; Phone: +6016-3346653; Email: mahendransekar@unikl.edu.my; Fax: +605-2536634

Authors

- Pei Teng Lum Department of Pharmaceutical Chemistry, Faculty of Pharmacy and Health Sciences, Universiti Kuala Lumpur Royal College of Medicine Perak, Ipoh 30450, Perak, Malaysia
- Siew Hua Gan School of Pharmacy, Monash University Malaysia, Bandar Sunway 47500, Selangor Darul Ehsan, Malaysia
- Srinivasa Reddy Bonam Institut National de la Santé et de la Recherche Médicale, Centre de Recherche des Cordeliers, Equipe-Immunopathologie et Immunointervention Thérapeutique, Sorbonne Université, Université de Paris, Paris 75006, France; orcid.org/0000-0002-2888-2418
- Mohd. Farooq Shaikh Neuropharmacology Research Strength, Jeffrey Cheah School of Medicine and Health Sciences, Monash University Malaysia, Bandar Sunway 47500, Selangor, Malaysia; Orcid.org/0000-0001-9865-6224

Complete contact information is available at: https://pubs.acs.org/10.1021/acschemneuro.0c00824

Author Contributions

P.T.L. and M.S. designed and conceived the ideas, collected the literature, interpreted the data, analyzed the data, and drafted and revised the manuscript. S.H.G., S.R.B., and M.F.S. analyzed the data and revised the manuscript. P.T.L., M.S., S.R.B., and M.F.S. created the figures. All authors read and approved the final manuscript.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors thank the Ministry of Higher Education (MOHE) Malaysia for the financial support provided via the Fundamental Research Grant Scheme [ref no. FRGS/1/ 2020/SKK06/UNIKL/02/4]. The authors also thank Universiti Kuala Lumpur Royal College of Medicine Perak, Malaysia, for providing necessary facilities and resources to complete this study for publication. The figures were created with the support of https://biorender.com under the paid subscription.

ABBREVIATIONS

3-NP, 3-nitropropionic acid; 5-HT, 5-hydroxytryptamine; AAV, adeno-associated virus; A2AR, adenosine A2A receptor; AC, adenyl cyclase; AChE, acetylcholinesterase; AMPK, AMPactivated protein kinase; ARE, antioxidant response element; ASO, antisense oligonucleotides; Atg, autophagy related; ATP, adenosine triphosphate; Bad, B-cell lymphoma 2 (Bcl-2)associated agonist of cell death; Bax, Bcl-2-associated X protein; BBB, blood-brain barrier; BDNF, brain-derived neurotrophic factor; CAG, cytosine-adenine-guanine; cAMP, cyclic adenosine monophosphate; CAT, catalase; CBP, cAMP-CREB-binding protein; CGN, cerebellar granule neuron; CNS, central nervous system; COX-2, cycloxygenase-2; CREB, cAMP response element binding protein; CRISPR, clustered regularly interspaced short palindromic repeats; CRISPR-Cas9, CRISPR-associated system 9; CS, citrate synthase; DA, dopamine; DARPP-32, dopamine- and cyclic-AMP-regulated phosphoprotein of molecular weight 32000; DDS, drug delivery systems; DMPK, drug metabolism and pharmacokinetics; Drp1, dynamin-1-like protein; ECGC, epigallocatechin gallate; ELT, escape latency; ERK, extracellular signalregulated kinase; ETC, electron transport chain; FDA, Food and Drug Administration; FGF, fibroblast growth factor; FST, forced swimming test; GABA, *γ*-aminobuytric acid; GCL, *γ*glutamyl-cysteinyl-ligase; GCS, γ -glutamylcysteine synthetase; GDNF, glial cell line-derived neurotrophic factor; GFAP, glial fibrillary acidic protein; GLP-1, glucagon-like protein; GFP, green fluorescent protein; GFPu, GFP-UPS reporter; GPx, glutathione peroxidase; GR, glutathione reductase; GSH, reduced glutathione; GSSG, glutathione disulfide; GST, glutathione S-transferase; GSTP1, glutathione S-transferase P1; HD, Huntington's disease; HO1, heme oxygenase-1; HSF1, heat shock factor 1; HSP, heat shock protein; HSP70, heat shock protein 70; Htt, huntingtin; ICR, Institute of Cancer Research; ig, intragastric gavage; ip, intraperitoneal injection; IL-1 β , interleukin-1 β ; IL-6, interleukin-6; iNOS, inducible nitric oxide synthase; JNK, c-Jun N-terminal kinase; KA, kynurenic acid; Keap1, Kelch-like ECH-associated protein 1; LC3-I, cytosolic form of LC3; LC3-II, LC3-phosphatidylethanolamine conjugate; LDH, lactate dehydrogenase; LKB1, liver kinase B1; LPA, lysophosphatidic acid; LPO, lipid

pubs.acs.org/chemneuro

peroxidation; MAO-A, monoamine oxidase A; MAPK, mitogen-activated protein kinase; MDA, malondialdehyde; MDH, malate dehydrogenase; MEF, mouse embryonic fibroblast; mGluR5, metabotropic glutamate receptor subtype 5; mHtt, mutant huntingtin; mMP, mitochondrial membrane potential; MMP2, matrix metalloproteinase-2; MMP9, matrix metalloproteinase-9; MMP, matrix metalloproteinase; MSN, medium spiny neurons; mTOR, mammalian target of rapamycin; MWM, Morris water maze test; N2a, neuro-2A; nAChR, nicotine acetylcholine receptor; NAD, nicotinamide adenine dinucleotide; NADH, NAD+ hydrogen (H); NE, norepinephrine; NF- κ B, nuclear factor kappa light chain enhancer of activated B cells; NGF, nerve growth factor; NIT, nitrite; NMDA, N-methyl-D-aspartate; NO, nitric oxide; NPSH, non-protein thiols; NQO1, NAD(P)H:quinone oxidoreductase 1; NR2B, NMDA receptor subtype 2B; Nrf2, nuclear factor erythroid 2-related factor 2; NBDD, nose to brain drug delivery; OFT, open field test; po, per os, oral administration; p53, tumor protein 53; p62, nucleoporin 62; PAINS, pan assay interference compounds; PC-12, pheochromocytoma-12; PKA, protein kinase; polyQ, polyglutamine; PPI, prepulse inhibition; PSD-95, postsynaptic density 95; PSH, protein thiols; PUR, puerarin; RNAi, RNA interference; RNS, reactive nitrogen species; ROS, reactive oxygen species; sc, subcutaneous injection; SD, Sprague-Dawley; SDH, succinate dehydrogenase; SIRT3, sirtuin-3; SNL, solanesol; SOD, superoxide dismutase; Sp1, specific protein-1; TATA, thymidine-adenine-thymidine-adenine; TBARS, thiobarbituric acid reactive substances; TBP, TATA-binding protein; TH, tyrosine hydroxylase; TIMP, tissue inhibitor of metalloproteinases; TL, transfer latency; TMP, tetramethylpyrazine; TNF- α , tumor necrosis factor α ; TSH, total thiols; TST, tail suspension test; TSTQ, time spent in target quadrant; UPS, ubiquitin proteasome system; VCM, vacuous chewing movement; WT, wild-type; ZFP, zinc finger motif protein

REFERENCES

(1) Ross, C. A., Aylward, E. H., Wild, E. J., Langbehn, D. R., Long, J. D., Warner, J. H., Scahill, R. I., Leavitt, B. R., Stout, J. C., Paulsen, J. S., et al. (2014) Huntington disease: natural history, biomarkers and prospects for therapeutics. *Nat. Rev. Neurol.* 10, 204–216.

(2) Gil, J. M., and Rego, A. C. (2008) Mechanisms of neurodegeneration in Huntington's disease. *Eur. J. Neurosci.* 27, 2803–2820.

(3) Bashir, H. (2019) Emerging therapies in Huntington's disease. Expert Rev. Neurother. 19, 983–995.

(4) Bates, G. P., Dorsey, R., Gusella, J. F., Hayden, M. R., Kay, C., Leavitt, B. R., Nance, M., Ross, C. A., Scahill, R. I., Wetzel, R., et al. (2015) Huntington disease. *Nat. Rev. Dis. Primers* 1, 15005.

(5) Bennett, E. J., Shaler, T. A., Woodman, B., Ryu, K.-Y., Zaitseva, T. S., Becker, C. H., Bates, G. P., Schulman, H., and Kopito, R. R. (2007) Global changes to the ubiquitin system in Huntington's disease. *Nature* 448, 704–708.

(6) Labbadia, J., and Morimoto, R. I. (2013) Huntington's disease: underlying molecular mechanisms and emerging concepts. *Trends Biochem. Sci.* 38, 378–385.

(7) Schaffar, G., Breuer, P., Boteva, R., Behrends, C., Tzvetkov, N., Strippel, N., Sakahira, H., Siegers, K., Hayer-Hartl, M., and Hartl, F. U. (2004) Cellular toxicity of polyglutamine expansion proteins: mechanism of transcription factor deactivation. *Mol. Cell* 15, 95–105. (8) Jana, N. R., Zemskov, E. A., Wang, G.-h., and Nukina, N. (2001) Altered proteasomal function due to the expression of polyglutamineexpanded truncated N-terminal huntingtin induces apoptosis by caspase activation through mitochondrial cytochrome c release. *Hum. Mol. Genet.* 10, 1049–1059. (9) Gunawardena, S., and Goldstein, L. S. (2005) Polyglutamine diseases and transport problems: deadly traffic jams on neuronal highways. *Arch. Neurol.* 62, 46–51.

(10) Liévens, J.-C., Woodman, B., Mahal, A., and Bates, G. P. (2002) Abnormal phosphorylation of synapsin I predicts a neuronal transmission impairment in the R6/2 Huntington's disease transgenic mice. *Mol. Cell. Neurosci.* 20, 638–648.

(11) Li, J.-Y., Plomann, M., and Brundin, P. (2003) Huntington's disease: a synaptopathy? *Trends Mol. Med.* 9, 414–420.

(12) Sun, Y., Savanenin, A., Reddy, P. H., and Liu, Y. F. (2001) Polyglutamine-expanded huntingtin promotes sensitization of Nmethyl-D-aspartate receptors via post-synaptic density 95. *J. Biol. Chem.* 276, 24713–24718.

(13) Almeida, S., Domingues, A., Rodrigues, L., Oliveira, C. R., and Rego, A. C. (2004) FK506 prevents mitochondrial-dependent apoptotic cell death induced by 3-nitropropionic acid in rat primary cortical cultures. *Neurobiol. Dis.* 17, 435–444.

(14) Choo, Y. S., Johnson, G. V., MacDonald, M., Detloff, P. J., and Lesort, M. (2004) Mutant huntingtin directly increases susceptibility of mitochondria to the calcium-induced permeability transition and cytochrome c release. *Hum. Mol. Genet.* 13, 1407–1420.

(15) Song, C., Zhang, Y., Parsons, C. G., and Liu, Y. F. (2003) Expression of polyglutamine-expanded huntingtin induces tyrosine phosphorylation of N-methyl-D-aspartate receptors. *J. Biol. Chem.* 278, 33364–33369.

(16) Tang, T.-S., Chen, X., Liu, J., and Bezprozvanny, I. (2007) Dopaminergic signaling and striatal neurodegeneration in Huntington's disease. *J. Neurosci.* 27, 7899–7910.

(17) Sandhir, R., Mehrotra, A., and Kamboj, S. S. (2010) Lycopene prevents 3-nitropropionic acid-induced mitochondrial oxidative stress and dysfunctions in nervous system. *Neurochem. Int.* 57, 579–587.

(18) Kulasekaran, G., and Ganapasam, S. (2015) Neuroprotective efficacy of naringin on 3-nitropropionic acid-induced mitochondrial dysfunction through the modulation of Nrf2 signaling pathway in PC12 cells. *Mol. Cell. Biochem.* 409, 199–211.

(19) Bae, B.-I., Xu, H., Igarashi, S., Fujimuro, M., Agrawal, N., Taya, Y., Hayward, S. D., Moran, T. H., Montell, C., Ross, C. A., et al. (2005) p53 mediates cellular dysfunction and behavioral abnormalities in Huntington's disease. *Neuron* 47, 29–41.

(20) Roos, R. A. (2010) Huntington's disease: a clinical review. Orphanet J. Rare Dis. 5, 40.

(21) Wheelock, V. L., Tempkin, T., Marder, K., Nance, M., Myers, R., Zhao, H., Kayson, E., Orme, C., Shoulson, I., and Huntington Study Group (2003) Predictors of nursing home placement in Huntington disease. *Neurology 60*, 998–1001.

(22) de Tommaso, M., Serpino, C., and Sciruicchio, V. (2011) Management of Huntington's disease: role of tetrabenazine. *Ther. Clin. Risk Manage.* 7, 123.

(23) Kumar, A., Kumar, V., Singh, K., Kumar, S., Kim, Y.-S., Lee, Y.-M., and Kim, J.-J. (2020) Therapeutic Advances for Huntington's Disease. *Brain Sci. 10*, 43.

(24) Wang, H., Chen, X., Li, Y., Tang, T.-S., and Bezprozvanny, I. (2010) Tetrabenazine is neuroprotective in Huntington's disease mice. *Mol. Neurodegener.* 5, 18.

(25) Coppen, E. M., and Roos, R. A. (2017) Current pharmacological approaches to reduce chorea in Huntington's disease. *Drugs* 77, 29–46.

(26) Rinaldi, C., and Wood, M. J. (2018) Antisense oligonucleotides: the next frontier for treatment of neurological disorders. *Nat. Rev. Neurol.* 14, 9–21.

(27) Garriga-Canut, M., Agustín-Pavón, C., Herrmann, F., Sánchez, A., Dierssen, M., Fillat, C., and Isalan, M. (2012) Synthetic zinc finger repressors reduce mutant huntingtin expression in the brain of R6/2 mice. *Proc. Natl. Acad. Sci. U. S. A. 109*, E3136–E3145.

(28) Crotti, A., and Glass, C. K. (2015) The choreography of neuroinflammation in Huntington's disease. *Trends Immunol.* 36, 364–373.

(29) Wang, Y., Dan, Y., Yang, D., Hu, Y., Zhang, L., Zhang, C., Zhu, H., Cui, Z., Li, M., and Liu, Y. (2014) The genus Anemarrhena Bunge: A review on ethnopharmacology, phytochemistry and pharmacology. J. Ethnopharmacol. 153, 42–60.

(30) Chen, J. K., Chen, T. T., and Crampton, L. (2004) *Chinese medical herbology and pharmacology*, Vol. 1267, Art of Medicine Press City of Industry, CA.

(31) Ji, D., Huang, Z.-Y., Fei, C.-H., Xue, W.-W., and Lu, T.-L. (2017) Comprehensive profiling and characterization of chemical constituents of rhizome of Anemarrhena asphodeloides Bge. *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 1060, 355–366.

(32) Jiang, W., Wei, W., Gaertig, M. A., Li, S., and Li, X.-J. (2015) Therapeutic effect of berberine on Huntington's disease transgenic mouse model. *PLoS One 10*, e0134142.

(33) Piwowar, A., Rembiałkowska, N., Rorbach-Dolata, A., Garbiec, A., Ślusarczyk, S., Dobosz, A., Długosz, A., Marchewka, Z., Matkowski, A., and Saczko, J. (2020) Anemarrhenae asphodeloides rhizoma Extract Enriched in Mangiferin Protects PC12 Cells against a Neurotoxic Agent-3-Nitropropionic Acid. *Int. J. Mol. Sci.* 21, 2510.

(34) Parvin, S., Easmin, D., Sheikh, A., Biswas, M., Sharma, S. C. D., Jahan, M. G. S., Islam, M. A., Shovon, M., and Roy, N. (2015) Nutritional analysis of date fruits (Phoenix dactylifera L.) in perspective of Bangladesh. *Am. J. Life Sci.* 3, 274–278.

(35) Al Alawi, R., Alhamdani, M. S. S., Hoheisel, J. D., and Baqi, Y. (2020) Antifibrotic and tumor microenvironment modulating effect of date palm fruit (*Phoenix dactylifera* L.) extracts in pancreatic cancer. *Biomed. Pharmacother.* 121, 109522.

(36) Essa, M. M., Singh, V., Guizani, N., Manivasagam, T., Thenmozhi, A. J., Bhat, A., Ray, B., and Chidambaram, S. B. (2019) *Phoenix dactylifera* L. Fruits Date Fruit Ameliorate Oxidative Stress in 3-NP Intoxicated PC12 Cells. *Int. J. Nutr. Pharmacol. Neurol. Dis.* 9, 41–47.

(37) Zhao, X., Feng, X., Peng, D., Liu, W., Sun, P., Li, G., Gu, L., and Song, J. L. (2016) Anticancer activities of alkaloids extracted from the Ba lotus seed in human nasopharyngeal carcinoma CNE-1 cells. *Exp. Ther. Med.* 12, 3113–3120.

(38) Abdel-Daim, M. M., Taha, R., Ghazy, E. W., and El-Sayed, Y. S. (2016) Synergistic ameliorative effects of sesame oil and alpha-lipoic acid against subacute diazinon toxicity in rats: hematological, biochemical, and antioxidant studies. *Can. J. Physiol. Pharmacol.* 94, 81–88.

(39) Im, A.-R., Chae, S.-W., Jun Zhang, G., and Lee, M.-Y. (2014) Neuroprotective effects of *Psoralea corylifolia* Linn seed extracts on mitochondrial dysfunction induced by 3-nitropropionic acid. *BMC Complementary Altern. Med.* 14, 370–377.

(40) Hmid, I., Elothmani, D., Hanine, H., Oukabli, A., and Mehinagic, E. (2017) Comparative study of phenolic compounds and their antioxidant attributes of eighteen pomegranate (Punica granatum L.) cultivars grown in Morocco. *Arabian J. Chem. 10*, S2675–S2684.

(41) Rathod, N., Biswas, D., Chitme, H., Ratna, S., Muchandi, I., and Chandra, R. (2012) Anti-urolithiatic effects of *Punica granatum* in male rats. *J. Ethnopharmacol.* 140, 234–238.

(42) BenSaad, L. A., Kim, K. H., Quah, C. C., Kim, W. R., and Shahimi, M. (2017) Anti-inflammatory potential of ellagic acid, gallic acid and punicalagin A&B isolated from Punica granatum. *BMC Complementary Altern. Med.* 17, 47.

(43) Al-Sabahi, B. N., Fatope, M. O., Essa, M. M., Subash, S., Al-Busafi, S. N., Al-Kusaibi, F. S., and Manivasagam, T. (2017) Pomegranate seed oil: Effect on 3-nitropropionic acid-induced neurotoxicity in PC12 cells and elucidation of unsaturated fatty acids composition. *Nutr. Neurosci.* 20, 40–48.

(44) Muley, B., Khadabadi, S., and Banarase, N. (2009) Phytochemical constituents and pharmacological activities of *Calendula officinalis* Linn (Asteraceae): a review. *Trop. J. Pharm. Res.* 8, 455–465.

(45) Shivasharan, B. D., Nagakannan, P., Thippeswamy, B. S., Veerapur, V. P., Bansal, P., and Unnikrishnan, M. K. (2013) Protective effect of *Calendula officinalis* Linn. flowers against 3-nitropropionic acid induced experimental Huntington's disease in rats. *Drug Chem. Toxicol.* 36, 466–473.

(46) Bhanumathy, M., Chandrasekar, S., Chandur, U., and Somasundaram, T. (2010) Phyto-pharmacology of Celastrus paniculatus: an Overview. *Int. J. Pharm. Sci. Drug Res.* 2, 176–181.

(47) Malik, J., Karan, M., and Dogra, R. (2017) Ameliorating effect of *Celastrus paniculatus* standardized extract and its fractions on 3nitropropionic acid induced neuronal damage in rats: Possible antioxidant mechanism. *Pharm. Biol.* 55, 980–990.

(48) Godkar, P. B., Gordon, R. K., Ravindran, A., and Doctor, B. P. (2004) *Celastrus paniculatus* seed water soluble extracts protect against glutamate toxicity in neuronal cultures from rat forebrain. *J. Ethnopharmacol.* 93, 213–219.

(49) Shinomol, G. K., and Muralidhara (2008) Prophylactic neuroprotective property of *Centella asiatica* against 3-nitropropionic acid induced oxidative stress and mitochondrial dysfunctions in brain regions of prepubertal mice. *NeuroToxicology* 29, 948–957.

(50) Shinomol, G. K., Ravikumar, H., and Muralidhara (2010) Prophylaxis with *Centella asiatica* confers protection to prepubertal mice against 3-nitropropionic-acid-induced oxidative stress in brain. *Phytother. Res.* 24, 885–892.

(51) Dhingra, D., and Valecha, R. (2007) Screening for antidepressant-like activity of *Convolvulus pluricaulis* Choisy in mice. *Pharmacol online* 1, 262–278.

(52) Malik, J., Choudhary, S., and Kumar, P. (2015) Protective effect of *Convolvulus pluricaulis* standardized extract and its fractions against 3-nitropropionic acid-induced neurotoxicity in rats. *Pharm. Biol. 53*, 1448–1457.

(53) Chunekar, K., and Pandey, G. (2002) Bhavaprakash Nighantu (Indian Materia Medica), Chaukhambha Bharati Academy, Varanasi.

(54) Kaur, M., Prakash, A., and Kalia, A. N. (2016) Neuroprotective potential of antioxidant potent fractions from *Convolvulus pluricaulis* Chois. in 3-nitropropionic acid challenged rats. *Nutr. Neurosci.* 19, 70–78.

(55) Prasad, P., Subhaktha, P., Narayana, A., and Rao, M. M. (2006) Medico-historical study of "asvattha" (sacred fig tree). *Bull. Indian Inst. Hist. Med. Hyderabad.* 36, 1–20.

(56) Aiyegoro, O., and Okoh, A. (2009) Use of bioactive plant products in combination with standard antibiotics: implications in antimicrobial chemotherapy. *J. Med. Plant Res.* 3, 1147–1152.

(57) Makhija, I. K., Sharma, I. P., and Khamar, D. (2010) Phytochemistry and Pharmacological properties of Ficus religiosa: an overview. *Ann. Biol. Res. 1*, 171–180.

(58) Bhangale, J. O., Acharya, N. S., and Acharya, S. R. (2016) Protective effect of *Ficus religiosa* (L.) against 3-nitropropionic acid induced Huntington disease. *Orient. Pharm. Exp. Med.* 16, 165–174.

(59) Choi, K. T. (2008) Botanical characteristics, pharmacological effects and medicinal components of Korean Panax ginseng CA Meyer. *Acta Pharmacol. Sin.* 29, 1109–1118.

(60) Jang, M., Lee, M. J., Kim, C. S., and Cho, I.-H. (2013) Korean red ginseng extract attenuates 3-nitropropionic acid-induced Huntington's-like symptoms. *Evid. Based Complement. Alternat. Med.* 2013, 1–17.

(61) Rosa, R. L. d., Nardi, G. M., Januário, A. G. d. F., Boçois, R., Bagatini, K. P., Bonatto, S. J. R., Pinto, A. d. O., Ferreira, J. R. N., Mariano, L. N. B., Niero, R., and Iagher, F. (2014) Anti-inflammatory, analgesic, and immunostimulatory effects of Luehea divaricata Mart. & Zucc.(Malvaceae) bark. *Braz. J. Pharm. Sci. 50*, 599–610.

(62) Tanaka, J. C. A., Silva, C. C. d., Dias Filho, B. P., Nakamura, C. V., Carvalho, J. E. d., and Foglio, M. A. (2005) Constituintes químicos de *Luehea divaricata* MART.(Tiliaceae). *Quim. Nova* 28, 834–837.

(63) Courtes, A. A., Arantes, L. P., Barcelos, R. P., da Silva, I. K., Boligon, A. A., Athayde, M. L., Puntel, R. L., and Soares, F. A. A. (2015) Protective effects of aqueous extract of *Luehea divaricata* against behavioral and oxidative changes induced by 3-nitropropionic acid in rats. *Evid. Based Complement. Alternat. Med.* 2015, 723431.

(64) Mancuso, C., and Santangelo, R. (2017) Panax ginseng and Panax quinquefolius: From pharmacology to toxicology. *Food Chem. Toxicol.* 107, 362–372. (65) Lian, X.-Y., Zhang, Z., and Stringer, J. L. (2005) Protective effects of ginseng components in a rodent model of neuro-degeneration. *Ann. Neurol.* 57, 642–648.

(66) Wu, J., Jeong, H. K., Bulin, S. E., Kwon, S. W., Park, J. H., and Bezprozvanny, I. (2009) Ginsenosides protect striatal neurons in a cellular model of Huntington's disease. *J. Neurosci. Res.* 87, 1904–1912.

(67) Tripathi, N., Shrivastava, D., Mir, B. A., Kumar, S., Govil, S., Vahedi, M., and Bisen, P. S. (2018) Metabolomic and biotechnological approaches to determine therapeutic potential of Withania somnifera (L.) Dunal: A review. *Phytomedicine* 50, 127–136.

(68) Rai, M., Jogee, P. S., Agarkar, G., and Santos, C. A. d. (2016) Anticancer activities of Withania somnifera: Current research, formulations, and future perspectives. *Pharm. Biol.* 54, 189–197.

(69) Kumar, P., and Kumar, A. (2009) Possible neuroprotective effect of *Withania somnifera* root extract against 3-nitropropionic acid-induced behavioral, biochemical, and mitochondrial dysfunction in an animal model of Huntington's disease. *J. Med. Food 12*, 591–600.

(70) Bartley, J. P., and Jacobs, A. L. (2000) Effects of drying on flavour compounds in Australian-grown ginger (Zingiber officinale). *J. Sci. Food Agric.* 80, 209–215.

(71) Sharma, M., Sharma, N., and Sharma, R. (2012) Neuroprotective effect of *Zingiber officinale* in 3-NP-induced huntington disease. *IOSR J. Pharm.* 2, 61–70.

(72) Stoilova, I., Krastanov, A., Stoyanova, A., Denev, P., and Gargova, S. (2007) Antioxidant activity of a ginger extract (Zingiber officinale). *Food Chem.* 102, 764–770.

(73) Shen, Y., Guan, Y., Song, X., He, J., Xie, Z., Zhang, Y., Zhang, H., and Tang, D. (2019) Polyphenols extract from lotus seedpod (*Nelumbo nucifera* Gaertn.): Phenolic compositions, antioxidant, and antiproliferative activities. *Food Sci. Nutr.* 7, 3062–3070.

(74) Wong, V., Wu, A., Wang, J., Liu, L., and Law, B. (2015) Neferine attenuates the protein level and toxicity of mutant huntingtin in PC-12 cells via induction of autophagy. *Molecules 20*, 3496–3514. (75) Cao, Q., Jiang, Y., Cui, S.-Y., Tu, P.-F., Chen, Y.-M., Ma, X.-L., Cui, X.-Y., Huang, Y.-L., Ding, H., Song, J.-Z., et al. (2016) Tenuifolin, a saponin derived from Radix Polygalae, exhibits sleepenhancing effects in mice. *Phytomedicine 23*, 1797–1805.

(76) Lee, C.-I., Han, J.-Y., Hong, J. T., and Oh, K.-W. (2013) 3, 4, 5-Trimethoxycinnamic acid (TMCA), one of the constituents of Polygalae Radix enhances pentobarbital-induced sleeping behaviors via GABA A ergic systems in mice. *Arch. Pharmacal Res.* 36, 1244– 1251.

(77) Wu, A.-G., Wong, V., Xu, S.-W., Chan, W.-K., Ng, C.-I., Liu, L., and Law, B. (2013) Onjisaponin B derived from Radix Polygalae enhances autophagy and accelerates the degradation of mutant α -synuclein and huntingtin in PC-12 cells. *Int. J. Mol. Sci.* 14, 22618–22641.

(78) Fu, J., Jin, J., Cichewicz, R. H., Hageman, S. A., Ellis, T. K., Xiang, L., Peng, Q., Jiang, M., Arbez, N., Hotaling, K., et al. (2012) trans-(-)-*e*-Viniferin increases mitochondrial sirtuin 3 (SIRT3), activates AMP-activated protein kinase (AMPK), and protects cells in models of Huntington Disease. *J. Biol. Chem.* 287, 24460–24472.

(79) Pedraza-Chaverrí, J., Reyes-Fermín, L. M., Nolasco-Amaya, E. G., Orozco-Ibarra, M., Medina-Campos, O. N., González-Cuahutencos, O., Rivero-Cruz, I., and Mata, R. (2009) ROS scavenging capacity and neuroprotective effect of α -mangostin against 3-nitropropionic acid in cerebellar granule neurons. *Exp. Toxicol.* Pathol. 61, 491–501.

(80) Cleren, C., Calingasan, N. Y., Chen, J., and Beal, M. F. (2005) Celastrol protects against MPTP-and 3-nitropropionic acid-induced neurotoxicity. *J. Neurochem.* 94, 995–1004.

(81) Westerheide, S. D., Bosman, J. D., Mbadugha, B. N., Kawahara, T. L., Matsumoto, G., Kim, S., Gu, W., Devlin, J. P., Silverman, R. B., and Morimoto, R. I. (2004) Celastrols as inducers of the heat shock response and cytoprotection. *J. Biol. Chem.* 279, 56053–56060.

(82) Liu, T. T., Zeng, Y., Tang, K., Chen, X., Zhang, W., and Xu, X. L. (2017) Dihydromyricetin ameliorates atherosclerosis in LDL receptor deficient mice. *Atherosclerosis 262*, 39–50.

(83) Le, L., Jiang, B., Wan, W., Zhai, W., Xu, L., Hu, K., and Xiao, P. (2016) Metabolomics reveals the protective of Dihydromyricetin on glucose homeostasis by enhancing insulin sensitivity. *Sci. Rep.* 6, 36184.

(84) Wang, J.-T., Jiao, P., Zhou, Y., and Liu, Q. (2016) Protective effect of dihydromyricetin against lipopolysaccharide-induced acute kidney injury in a rat model. *Med. Sci. Monit.* 22, 454.

(85) Mu, S., Li, Y., Liu, B., Wang, W., Chen, S., Wu, J., OuYang, L., Zhu, Y., Li, K., Zhan, M., et al. (2016) Dihydromyricetin ameliorates 3NP-induced behavioral deficits and striatal injury in rats. *J. Mol. Neurosci.* 60, 267–275.

(86) Durg, S., Veerapur, V. P., Neelima, S., and Dhadde, S. B. (2017) Antidiabetic activity of Embelia ribes, embelin and its derivatives: A systematic review and meta-analysis. *Biomed. Pharmacother.* 86, 195– 204.

(87) Mahendran, S., Badami, S., and Maithili, V. (2011) Evaluation of antidiabetic effect of embelin from Embelia ribes in alloxan induced diabetes in rats. *Biomed. Prev. Nutr.* 1, 25–31.

(88) Dhadde, S. B., Nagakannan, P., Roopesh, M., Kumar, S. A., Thippeswamy, B., Veerapur, V. P., and Badami, S. (2016) Effect of embelin against 3-nitropropionic acid-induced Huntington's disease in rats. *Biomed. Pharmacother.* 77, 52–58.

(89) Zhang, L., Ho, C. T., Zhou, J., Santos, J. S., Armstrong, L., and Granato, D. (2019) Chemistry and biological activities of processed Camellia sinensis teas: A comprehensive review. *Compr. Rev. Food Sci. Food Saf.* 18, 1474–1495.

(90) Kumar, P., and Kumar, A. (2009) Protective effects of epigallocatechin gallate following 3-nitropropionic acid-induced brain damage: possible nitric oxide mechanisms. *Psychopharmacology* 207, 257–270.

(91) Matos, M. J., Santana, L., Uriarte, E., Abreu, O. A., Molina, E., and Yordi, E. G. (2015) Coumarins - An important class of phytochemicals. *In Phytochemicals-Isolation, Characterisation and Role in Human Health*, 113–140.

(92) Karandikar, A., and Thangarajan, S. (2017) Protective activity of Esculetin against 3-nitropropionic acid induced neurotoxicity via scavenging reactive oxygen species in male wistar rats. *Int. J. Pharmacogn. Phytochem. Res.* 9, 722–732.

(93) Kavitha, C., Rajamani, K., and Vadivel, E. (2010) Coleus forskohlii A comprehensive review on morphology, phytochemistry and pharmacological aspects. *J. Med. Plant Res.* 4, 278–285.

(94) Mehan, S., Parveen, S., and Kalra, S. (2017) Adenyl cyclase activator forskolin protects against Huntington's disease-like neuro-degenerative disorders. *Neural Regener. Res.* 12, 290–300.

(95) Fontán-Lozano, Á., Romero-Granados, R., Pérez-Villegas, E. M., and Carrión, Á. M. (2012) The role of CREB in neuronal plasticity, learning and memory, and in neuropsychiatric disorders, *Transcription Factors CREB and NF-κB: Involvement in Synaptic Plasticity and Memory Formation*, pp 22–42, Bentham science, Canada.

(96) Malla, A., and Ramalingam, S. (2018) Health Perspectives of an Isoflavonoid Genistein and its Quantification in Economically Important Plants, in *Role of Materials Science in Food Bioengineering* (Grumezescu, A. M., and Holban, A. M., Eds.), Chapter 11, pp 353–379, Academic Press.

(97) Menze, E. T., Esmat, A., Tadros, M. G., Abdel-Naim, A. B., and Khalifa, A. E. (2015) Genistein improves 3-NPA-induced memory impairment in ovariectomized rats: Impact of its antioxidant, antiinflammatory and acetylcholinesterase modulatory properties. *PLoS One 10*, e0117223.

(98) Barnes, S., Kim, H., Darley-Usmar, V., Patel, R., Xu, J., Boersma, B., and Luo, M. (2000) Beyond ER α and ER β : estrogen receptor binding is only part of the isoflavone story. *J. Nutr. 130*, 6568–657S.

(99) Steiner, B. M., McClements, D. J., and Davidov-Pardo, G. (2018) Encapsulation systems for lutein: A review. *Trends Food Sci. Technol.* 82, 71–81.

(100) Binawade, Y., and Jagtap, A. (2013) Neuroprotective effect of lutein against 3-nitropropionic acid-induced Huntington's disease-like

symptoms: Possible behavioral, biochemical, and cellular alterations. *J. Med. Food 16*, 934–943.

(101) Tariq, M., Khan, H. A., Elfaki, I., Al Deeb, S., and Al Moutaery, K. (2005) Neuroprotective effect of nicotine against 3nitropropionic acid (3-NP)-induced experimental Huntington's disease in rats. *Brain Res. Bull.* 67, 161–168.

(102) O'neill, M., Murray, T., Lakics, V., Visanji, N., and Duty, S. (2002) The role of neuronal nicotinic acetylcholine receptors in acute and chronic neurodegeneration. *Curr. Drug Targets: CNS Neurol. Disord.* 1, 399–411.

(103) Ravikumar, R., Fugaccia, I., Scheff, S. W., Geddes, J. W., Srinivasan, C., and Toborek, M. (2005) Nicotine attenuates morphological deficits in a contusion model of spinal cord injury. *J. Neurotrauma* 22, 240–251.

(104) Túnez, I., Montilla, P., Muñoz, M. C., and Drucker-Colín, R. (2004) Effect of nicotine on 3-nitropropionic acid-induced oxidative stress in synaptosomes. *Eur. J. Pharmacol.* 504, 169–175.

(105) Cormier, A., Morin, C., Zini, R., Tillement, J.-P., and Lagrue, G. (2001) In vitro effects of nicotine on mitochondrial respiration and superoxide anion generation. *Brain Res.* 900, 72–79.

(106) Song, Y., Jing, W., Yan, R., and Wang, Y. (2015) Research progress of the studies on the roots of Peucedanum praeruptorum dunn (Peucedani radix). *Pak. J. Pharm. Sci.* 28, 71–81.

(107) Wang, L., Wang, J., Yang, L., Zhou, S.-M., Guan, S.-Y., Yang, L.-K., Shi, Q.-X., Zhao, M.-G., and Yang, Q. (2017) Effect of Praeruptorin C on 3-nitropropionic acid induced Huntington's disease-like symptoms in mice. *Biomed. Pharmacother.* 86, 81–87.

(108) Shi, W., Yang, L., Shi, Q., Yang, Y., Zhou, S., et al. (2015) The Anti-Depressant Effect of Praeruptorin C on the Chronic Unpredictable Mild Stress Mouse Model. *Clin. Exp. Pharmacol.* 5, 1000195.

(109) Liu, Z., Qi, Y., Cheng, Z., Zhu, X., Fan, C., and Yu, S. (2016) The effects of ginsenoside Rg1 on chronic stress induced depressionlike behaviors, BDNF expression and the phosphorylation of PKA and CREB in rats. *Neuroscience* 322, 358–369.

(110) Lim, D., Lee, C., Kim, I.-H., and Kim, Y. (2013) Antiinflammatory effects of total isoflavones from Pueraria lobata on cerebral ischemia in rats. *Molecules* 18, 10404–10412.

(111) Zhang, Z., Lam, T. N., and Zuo, Z. (2013) Radix Puerariae: an overview of its chemistry, pharmacology, pharmacokinetics, and clinical use. *J. Clin. Pharmacol.* 53, 787–811.

(112) Mahdy, H. M., Mohamed, M. R., Emam, M. A., Karim, A. M., Abdel-Naim, A., and Khalifa, A. E. (2014) The anti-apoptotic and anti-inflammatory properties of puerarin attenuate 3-nitropropionic-acid induced neurotoxicity in rats. *Can. J. Physiol. Pharmacol.* 92, 252–258.

(113) Mahdy, H. M., Mohamed, M. R., Emam, M. A., Karim, A. M., Abdel-Naim, A. B., and Khalifa, A. E. (2014) Puerarin ameliorates 3nitropropionic acid-induced neurotoxicity in rats: possible neuromodulation and antioxidant mechanisms. *Neurochem. Res.* 39, 321– 332.

(114) Lesjak, M., Beara, I., Simin, N., Pintać, D., Majkić, T., Bekvalac, K., Orčić, D., and Mimica-Dukić, N. (2018) Antioxidant and anti-inflammatory activities of quercetin and its derivatives. *J. Funct. Foods* 40, 68–75.

(115) Serban, M. C., Sahebkar, A., Zanchetti, A., Mikhailidis, D. P., Howard, G., Antal, D., Andrica, F., Ahmed, A., Aronow, W. S., Muntner, P., et al. (2016) Effects of quercetin on blood pressure: a systematic review and meta-analysis of randomized controlled trials. *J. Am. Heart Assoc. 5*, No. e002713.

(116) Chakraborty, J., Singh, R., Dutta, D., Naskar, A., Rajamma, U., and Mohanakumar, K. P. (2014) Quercetin improves behavioral deficiencies, restores astrocytes and microglia, and reduces serotonin metabolism in 3-nitropropionic acid-induced rat model of Huntington's disease. *CNS Neurosci. Ther.* 20, 10–19.

(117) Berman, A. Y., Motechin, R. A., Wiesenfeld, M. Y., and Holz, M. K. (2017) The therapeutic potential of resveratrol: a review of clinical trials. *NPJ. Precis. Oncol.* 1, 35.

(118) Kumar, P., Padi, S. S. V., Naidu, P. S., and Kumar, A. (2006) Effect of resveratrol on 3-nitropropionic acid-induced biochemical

and behavioural changes: possible neuroprotective mechanisms. *Behav. Pharmacol.* 17, 485–492.

(119) Uddandrao, V. S., Brahmanaidu, P., and Saravanan, G. (2018) Therapeutical perspectives of S-allylcysteine: effect on diabetes and other disorders in animal models. *Cardiovasc. Hematol. Agents Med. Chem.* 15, 71–77.

(120) Elinos-Calderón, D., Robledo-Arratia, Y., Pérez-De La Cruz, V., Maldonado, P. D., Galván-Arzate, S., Pedraza-Chaverrí, J., and Santamaría, A. (2010) Antioxidant strategy to rescue synaptosomes from oxidative damage and energy failure in neurotoxic models in rats: Protective role of S-allylcysteine. *J. Neural Transm.* 117, 35–44. (121) Herrera-Mundo, M. N., Silva-Adaya, D., Maldonado, P. D., Galván-Arzate, S., Andrés-Martínez, L., Pérez-De La Cruz, V., Pedraza-Chaverrí, J., and Santamaría, A. (2006) S-Allylcysteine prevents the rat from 3-nitropropionic acid-induced hyperactivity, early markers of oxidative stress and mitochondrial dysfunction. *Neurosci. Res.* 56, 39–44.

(122) Miraj, S., and Kiani, S. (2016) Bioactivity of Sesamum indicum: A review study. *Der Pharm. Lett.* 8, 328–334.

(123) Kumar, P., Kalonia, H., and Kumar, A. (2010) Protective effect of sesamol against 3-nitropropionic acid-induced cognitive dysfunction and altered glutathione redox balance in rats. *Basic Clin. Pharmacol. Toxicol.* 107, 577–582.

(124) Kumar, P., Kalonia, H., and Kumar, A. (2009) Sesamol attenuate 3-nitropropionic acid-induced Huntington-like behavioral, biochemical, and cellular alterations in rats. *J. Asian Nat. Prod. Res.* 11, 439–450.

(125) Kishore, K. (2014) Monograph of tobacco (*Nicotiana tabacum*). Indian J. Drugs 2, 5–23.

(126) Mehan, S., Monga, V., Rani, M., Dudi, R., and Ghimire, K. (2018) Neuroprotective effect of solanesol against 3-nitropropionic acid-induced Huntington's disease-like behavioral, biochemical, and cellular alterations: Restoration of coenzyme-Q10-mediated mitochondrial dysfunction. *Indian J. Pharmacol.* 50, 309.

(127) Choi, Y. H., and Park, H. Y. (2012) Anti-inflammatory effects of spermidine in lipopolysaccharide-stimulated BV2 microglial cells. *J. Biomed. Sci.* 19, 31.

(128) Jamwal, S., and Kumar, P. (2016) Spermidine ameliorates 3nitropropionic acid (3-NP)-induced striatal toxicity: Possible role of oxidative stress, neuroinflammation, and neurotransmitters. *Physiol. Behav.* 155, 180–187.

(129) Wei, Z., Dong, C., Guan, L., Wang, Y., Huang, J., and Wen, X. (2020) A metabolic exploration of the protective effect of *Ligusticum* wallichii on IL-1 β -injured mouse chondrocytes. *Chin. Med.* 15, 12.

(130) Danduga, R. C. S. R., Dondapati, S. R., Kola, P. K., Grace, L., Tadigiri, R. V. B., and Kanakaraju, V. K. (2018) Neuroprotective activity of tetramethylpyrazine against 3-nitropropionic acid induced Huntington's disease-like symptoms in rats. *Biomed. Pharmacother. 105*, 1254–1268.

(131) Yu, L., Jiang, X., Zhang, Y., Liao, M., Ma, R., and Yu, T. (2012) Antidepressant-like activity of Tetramethylpyrazine measured by chronic experimental method in rat model of depression. *Pharmacol. Pharm.* 3, 52–57.

(132) Jiang, B., Huang, C., Chen, X.-F., Tong, L.-J., and Zhang, W. (2015) Tetramethylpyrazine produces antidepressant-like effects in mice through promotion of BDNF signaling pathway. *Int. J. Neuropsychopharmacol.* 18, pyv010.

(133) Peng, L., Song, X., Shi, X., Li, J., and Ye, C. (2008) An improved HPLC method for simultaneous determination of phenolic compounds, purine alkaloids and theanine in Camellia species. *J. Food Compos. Anal.* 21, 559–563.

(134) Thangarajan, S., Deivasigamani, A., Natarajan, S. S., Krishnan, P., and Mohanan, S. K. (2014) Neuroprotective activity of L-theanine on 3-nitropropionic acid-induced neurotoxicity in rat striatum. *Int. J. Neurosci.* 124, 673–684.

(135) Kim, T. I., Lee, Y. K., Park, S. G., Choi, I. S., Ban, J. O., Park, H. K., Nam, S.-Y., Yun, Y. W., Han, S. B., Oh, K. W., and Hong, J. T. (2009) l-Theanine, an amino acid in green tea, attenuates β -amyloid-induced cognitive dysfunction and neurotoxicity: reduction in

oxidative damage and inactivation of ERK/p38 kinase and NF- κ B pathways. *Free Radical Biol. Med.* 47, 1601–1610.

(136) Cho, Y.-J., Choi, S.-H., Lee, R., Hwang, H., Rhim, H., Cho, I.-H., Kim, H.-C., Lee, J.-I., Hwang, S.-H., and Nah, S.-Y. (2020) Ginseng Gintonin Contains Ligands for GPR40 and GPR55. *Molecules* 25, 1102.

(137) Hwang, S. H., Lee, B.-H., Kim, H.-J., Cho, H.-J., Shin, H.-C., Im, K.-S., Choi, S.-H., Shin, T.-J., Lee, S.-M., Nam, S. W., et al. (2013) Suppression of metastasis of intravenously-inoculated B16/F10 melanoma cells by the novel ginseng-derived ingredient, gintonin: involvement of autotaxin inhibition. *Int. J. Oncol.* 42, 317–326.

(138) Kim, S., Kim, M.-S., Park, K., Kim, H.-J., Jung, S.-W., Nah, S.-Y., Han, J.-S., and Chung, C. (2016) Hippocampus-dependent cognitive enhancement induced by systemic gintonin administration. *J. Ginseng Res.* 40, 55–61.

(139) Jang, M., Choi, J. H., Chang, Y., Lee, S. J., Nah, S.-Y., and Cho, I.-H. (2019) Gintonin, a ginseng-derived ingredient, as a novel therapeutic strategy for Huntington's disease: activation of the Nrf2 pathway through lysophosphatidic acid receptors. *Brain, Behav., Immun.* 80, 146–162.

(140) Gopinath, K., Prakash, D., and Sudhandiran, G. (2011) Neuroprotective effect of naringin, a dietary flavonoid against 3nitropropionic acid-induced neuronal apoptosis. *Neurochem. Int. 59*, 1066–1073.

(141) Gopinath, K., and Sudhandiran, G. (2016) Protective effect of naringin on 3-nitropropionic acid-induced neurodegeneration through the modulation of matrix metalloproteinases and glial fibrillary acidic protein. *Can. J. Physiol. Pharmacol. 94*, 65–71.

(142) Gopinath, K., and Sudhandiran, G. (2012) Naringin modulates oxidative stress and inflammation in 3-nitropropionic acid-induced neurodegeneration through the activation of nuclear factor-erythroid 2-related factor-2 signalling pathway. *Neuroscience* 227, 134–143.

(143) Gao, Y., Chu, S.-F., Li, J.-P., Zhang, Z., Yan, J.-Q., Wen, Z.-L., Xia, C.-Y., Mou, Z., Wang, Z.-Z., He, W.-B., Guo, X.-F., Wei, G.-N., and Chen, N.-H. (2015) Protopanaxatriol protects against 3-nitropropionic acid-induced oxidative stress in a rat model of Huntington's disease. *Acta Pharmacol. Sin. 36*, 311–322.

(144) Liu, Y., Hettinger, C. L., Zhang, D., Rezvani, K., Wang, X., and Wang, H. (2014) Sulforaphane enhances proteasomal and autophagic activities in mice and is a potential therapeutic reagent for Huntington's disease. *J. Neurochem.* 129, 539–547.

(145) Jang, M., and Cho, I.-H. (2016) Sulforaphane ameliorates 3nitropropionic acid-induced striatal toxicity by activating the Keap1-Nrf2-ARE pathway and inhibiting the MAPKs and NF-*x*B pathways. *Mol. Neurobiol.* 53, 2619–2635.

(146) Huang, C.-L., Yang, J.-M., Wang, K.-C., Lee, Y.-C., Lin, Y.-L., Yang, Y.-C., and Huang, N.-K. (2011) *Gastrodia elata* prevents huntingtin aggregations through activation of the adenosine A2A receptor and ubiquitin proteasome system. *J. Ethnopharmacol.* 138, 162–168.

(147) Huang, N., Lin, J., Lin, J., Lin, C., Liu, E., et al. (2011) A new drug design targeting the adenosinergic system for Huntington's disease. *PLoS One 6*, e20934.

(148) Pohl, F., and Kong Thoo Lin, P. (2018) The potential use of plant natural products and plant extracts with antioxidant properties for the prevention/treatment of neurodegenerative diseases: in vitro, in vivo and clinical trials. *Molecules* 23, 3283.

(149) Brand, M. D. (2010) The sites and topology of mitochondrial superoxide production. *Exp. Gerontol.* 45, 466–472.

(150) Feng, S.-T., Wang, Z.-Z., Yuan, Y.-H., Sun, H.-M., Chen, N.-H., and Zhang, Y. (2019) Mangiferin: a multipotent natural product preventing neurodegeneration in Alzheimer's and Parkinson's disease models. *Pharmacol. Res.* 146, 104336.

(151) Singh, S., Jamwal, S., and Kumar, P. (2015) Piperine enhances the protective effect of curcumin against 3-NP induced neurotoxicity: possible neurotransmitters modulation mechanism. *Neurochem. Res.* 40, 1758–1766. (152) Niu, X., Chen, J., and Gao, J. (2019) Nanocarriers as a powerful vehicle to overcome blood-brain barrier in treating neurodegenerative diseases: Focus on recent advances. *Asian J. Pharm. Sci.* 14, 480–496.

(153) Poovaiah, N., Davoudi, Z., Peng, H., Schlichtmann, B., Mallapragada, S., Narasimhan, B., and Wang, Q. (2018) Treatment of neurodegenerative disorders through the blood-brain barrier using nanocarriers. *Nanoscale 10*, 16962–16983.

(154) Rahaman, S. T. (2019) The role of nanomedicine in the treatment of neurodegenerative disorders, *Nanobiotechnology in Neurodegenerative Diseases*, pp 49–63, Springer.

(155) Saraiva, C., Praça, C., Ferreira, R., Santos, T., Ferreira, L., and Bernardino, L. (2016) Nanoparticle-mediated brain drug delivery: overcoming blood-brain barrier to treat neurodegenerative diseases. *J. Controlled Release* 235, 34–47.

(156) Teleanu, D. M., Negut, I., Grumezescu, V., Grumezescu, A. M., and Teleanu, R. I. (2019) Nanomaterials for drug delivery to the central nervous system. *Nanomaterials 9*, 371.

(157) Ramalho, M. J., and Pereira, M. C. (2016) Preparation and characterization of polymeric nanoparticles: an interdisciplinary experiment. *J. Chem. Educ.* 93, 1446–1451.

(158) Long, Y., Yang, Q., Xiang, Y., Zhang, Y., Wan, J., Liu, S., Li, N., and Peng, W. (2020) Nose to brain drug delivery-a promising strategy for active components from herbal medicine for treating cerebral ischemia reperfusion. *Pharmacol. Res.* 159, 104795.

(159) Kumari, A., Singla, R., Guliani, A., and Yadav, S. K. (2014) Nanoencapsulation for drug delivery. *EXCLI journal 13*, 265.

(160) Rafiee, Z., Nejatian, M., Daeihamed, M., and Jafari, S. M. (2019) Application of different nanocarriers for encapsulation of curcumin. *Crit. Rev. Food Sci. Nutr.* 59, 3468–3497.

(161) Xu, W., Ling, P., and Zhang, T. (2013) Polymeric micelles, a promising drug delivery system to enhance bioavailability of poorly water-soluble drugs. *J. Drug Delivery 2013*, 340315.

(162) Shelley, H., and Babu, R. J. (2018) Role of cyclodextrins in nanoparticle-based drug delivery systems. *J. Pharm. Sci.* 107, 1741–1753.

(163) Patel, V. R., and Agrawal, Y. (2011) Nanosuspension: An approach to enhance solubility of drugs. *J. Adv. Pharm. Technol. Res. 2*, 81.

(164) Yen, C.-C., Chen, Y.-C., Wu, M.-T., Wang, C.-C., and Wu, Y.-T. (2018) Nanoemulsion as a strategy for improving the oral bioavailability and anti-inflammatory activity of and rographolide. *Int. J. Nanomed.* 13, 669.

(165) Lipinski, C. A., Lombardo, F., Dominy, B. W., and Feeney, P. J. (1997) Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv. Drug Delivery Rev.* 23, 3–25.

(166) Veber, D. F., Johnson, S. R., Cheng, H.-Y., Smith, B. R., Ward, K. W., and Kopple, K. D. (2002) Molecular properties that influence the oral bioavailability of drug candidates. *J. Med. Chem.* 45, 2615–2623.

(167) Baell, J. B. (2016) Feeling nature's PAINS: natural products, natural product drugs, and pan assay interference compounds (PAINS). J. Nat. Prod. 79, 616–628.

(168) Baell, J. B., and Holloway, G. A. (2010) New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays. *J. Med. Chem.* 53, 2719–2740.